

A new species of *Rimicaris* (Crustacea: Decapoda: Caridea: Alvinocarididae) from hydrothermal vent fields on the Mid-Cayman Spreading Centre, Caribbean

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Rimicaris hybisae sp. nov. is described from hydrothermal vent fields on the world's deepest seafloor spreading centre, the Mid-Cayman Spreading Centre (MCSC), Caribbean, at depths of 2300–4960 m. The new species is described and illustrated on the basis of 17 specimens. Brief notes on the distribution and habitat of the new species are provided. Molecular phylogenetic data from mitochondrial COI (460 base pair (bp)), 16S ribosomal RNA (549 bp) and nuclear 18S ribosomal RNA (576 bp) regions is used to complement the description. Morphological variation within *R. hybisae* sp. nov. and morphological affinities with previously described species are discussed. Based on morphological and molecular evidence, the new species is provisionally assigned to the genus *Rimicaris*, and differs from all known species in the genus by a distinctive pair of 'pores' on the posterior lobes of its four-lobed dorsal organ. An emended diagnosis for *Rimicaris* is provided. *Rimicaris hybisae* sp. nov. is the first taxon to be described from MCSC vent fields. This record extends the known geographical range of *Rimicaris* into the Caribbean Sea and constitutes the deepest documented occurrence of alvinocaridid shrimp.

Keywords: Crustacea, Decapoda, Caridea, Alvinocarididae, *Rimicaris*, new species, Cayman, hydrothermal vents

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INTRODUCTION

All known species of the caridean family Alvinocarididae Christoffersen, 1986 occur exclusively in hydrothermal vents or cold (brine and hydrocarbon) seeps (see Table 1 and references therein). Alvinocaridids are known from a considerable number of vents and seeps in the Atlantic, Pacific and Indian Oceans (Table 1). Alvinocaridid shrimp are the dominant macrofaunal invertebrates at several vents along the Mid-Atlantic Ridge (MAR) (e.g. Van Dover *et al.*, 1988; Segonzac, 1992; Gebruk *et al.*, 1993, 1997; Van Dover, 2000; Martin & Shank, 2005) and at known vent fields in the Indian Ocean (Hashimoto *et al.*, 2001; Van Dover *et al.*, 2001; Watabe & Hashimoto, 2002). The Alvinocarididae presently comprises 26 described species in 8 genera, all from chemosynthetic environments in the bathymetric range 252–4088 m (Table 1).

Alvinocaris Williams & Chace, 1982 is presently the only genus known to inhabit both hydrothermal vents and cold seeps (Table 1). *Alvinocaris*, *Rimicaris* Williams & Rona, 1986, *Chorocaris* Martin & Hessler, 1990 and *Opaepele* Williams & Dobbs, 1995 were originally assigned to the caridean family Bresiliidae Calman, 1896 (see Martin &

Davis, 2001; Komai & Segonzac, 2003), whereas the family Mirocarididae Vereshchaka, 1997 was established to accommodate the genus *Mirocaris* Vereshchaka, 1997. Komai & Segonzac (2003) subsequently assigned all these genera to the Alvinocarididae, and synonymized the family Mirocarididae with the Alvinocarididae.

The family Bresiliidae is now only represented by two genera (De Grave *et al.*, 2009), *Bresilia* Calman, 1896 and the monotypic *Encantada* Wicksten, 1989, neither of which are known to occur in chemosynthetic habitats (Wicksten, 1989; Komai & Yamada, 2010, 2011). The family Alvinocarididae (Christoffersen, 1986, 1990; Segonzac *et al.*, 1993; Komai & Segonzac, 2003) is now regarded as a valid monophyletic family (e.g. Komai & Segonzac, 2005, 2008; Martin & Haney, 2005; De Grave *et al.*, 2009), morphologically distinct from opportunistic shrimp species recorded from vents and seeps (see Martin & Haney, 2005; Desbruyères *et al.*, 2006 for recent reviews).

Recently, two high-temperature hydrothermal vent fields and chemosynthetic communities were discovered on the world's deepest seafloor spreading centre, the Mid-Cayman Spreading Centre (MCSC), Caribbean (Connelly *et al.*, in press). The ~110 km long, ultraslow-spreading (15 mm yr⁻¹) MCSC has been active for approximately 49 My (Rosencrantz *et al.*, 1988; German *et al.*, 2010) and is located in a deep trough, geographically and tectonically isolated from the global mid-ocean ridge system (Ballard *et al.*, 1979).

The Beebe Vent Field (BVF) consists of a sulphide mound (80 m diameter, 50 m height) surmounted with several actively

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Table 1. Summary of known geographical distribution, bathymetric range and habitat of alvinocaridid shrimp species (confirmed locations and fully described species only).

Species	Site(s)	Depth (m)	Habitat	Primary references
<i>Alvinocaridinides formosa</i>	North-east Taiwan: Gueishandao	252–275	Vent	Komai & Chan, 2010
<i>Alvinocaris alexander</i>	KR: Rumble V Seamount, Brothers Caldera	367–1346	Vent	Ahyong, 2009
<i>Alvinocaris brevitelsonis</i>	OT: Minami–Ensei Knoll	705	Vent	Kikuchi & Hashimoto, 2000 (see also Komai & Segonzac, 2005)
<i>Alvinocaris chelys</i>	North-east Taiwan: Gueishandao	252–300	Vent	Komai & Chan, 2010
<i>Alvinocaris dissimilis</i>	OT: Minami–Ensei Knoll	705	Vent	Komai & Segonzac, 2005
<i>Alvinocaris komai</i>	ELSC: ABE, Kilo Moana, TowCam, Tu'i Malila	1880–2700	Vent	Zelnio & Hourdez, 2009
<i>Alvinocaris longirostris</i>	OT: Iheya Ridge, Hatoma Knoll; SB: Off Hatsushima site	1053–1627	Vent and seep	Kikuchi & Ohta, 1995; Fujikura <i>et al.</i> , 1995; Watabe & Miyake, 2000; Ohta & Kim, 2001
<i>Alvinocaris lusca</i>	GR: Rose Garden; EPR: 9°N	2450–2520	Vent	Williams & Chace, 1982; Shank <i>et al.</i> , 1999
<i>Alvinocaris markensis</i>	MAR: Lucky Strike; Rainbow; Broken Spur; TAG; Snake Pit; Logatchev	1693–3650	Vent	Williams, 1988; Shank <i>et al.</i> , 1999
<i>Alvinocaris methanophila</i>	Blake Ridge Diapir	2155	Seep	Komai <i>et al.</i> , 2005
<i>Alvinocaris muricola</i>	GoM: Florida Escarpment; Barbados Accretionary prism; West African equatorial margin, Congo Basin; Blake Ridge Diapir	1697–3277	Seep	Williams, 1988; Komai & Segonzac, 2005; Komai <i>et al.</i> , 2005
<i>Alvinocaris niwa</i>	KR: Rumble V Seamount, Brothers Caldera	360–1538	Vent	Webber, 2004
<i>Alvinocaris stactophila</i>	GoM: Louisiana Slope	534	Seep	Williams, 1988
<i>Alvinocaris williamsi</i>	MAR: Menez Gwen	850–865	Vent	Shank & Martin, 2003
<i>Chorocaris chacei</i>	MAR: Moytirra; Lucky Strike; TAG; Snake Pit; Logatchev	1600–3650	Vent	Williams & Rona, 1986; Komai & Segonzac, 2008; Copley <i>et al.</i> , in press
<i>Chorocaris paulexa</i>	EPR: 17°–21°S	2573–2832	Vent	Martin & Shank, 2005
<i>Chorocaris vandoverae</i>	MBAB: Alice Springs, Burke	3640–3660	Vent	Martin & Hessler, 1990
<i>Mirocaris fortunata</i>	MAR: Moytirra; Menez Gwen; Lucky Strike; Rainbow; Broken Spur; TAG; Snake Pit, Logatchev; Ashadze; Turtle Pits	850–3480	Vent	Martin & Christiansen, 1995; Vereshchaka, 1997; Shank <i>et al.</i> , 1999; Komai & Segonzac, 2003; Komai <i>et al.</i> , 2007; Fabri <i>et al.</i> , 2011; Copley <i>et al.</i> , 2011
<i>Mirocaris indica</i>	CIR: Kairei, Edmond	2422–3300	Vent	Komai <i>et al.</i> , 2006
<i>Nautilicaris saintlaurenti</i>	NFB: White Lady, Mussel Valley; LB: Hine Hina, Vai Lili; KR: Brothers Caldera	1604–2000	Vent	Komai & Segonzac, 2004; Ahyong, 2009
<i>Opaepele loihi</i>	Hawaii: Loihi Seamount	980	Vent	Williams & Dobbs, 1995
<i>Opaepele susanna</i>	MAR: Sisters Peak; Lilliput; Semenov	1500–2986	Vent	Komai <i>et al.</i> , 2007; Beltenev <i>et al.</i> , 2009
<i>Opaepele vavilovi</i>	MAR: Broken Spur	~3090	Vent	Lunina & Vereshchaka, 2010
<i>Rimicaris exoculata</i>	MAR: Moytirra; Rainbow; Lucky Strike; Broken Spur; TAG; Snake Pit; Logatchev; Ashadze; Mephisto	1700–4088	Vent	Williams & Rona, 1986; Komai <i>et al.</i> , 2007; Komai & Segonzac, 2008; Copley <i>et al.</i> , in press
<i>Rimicaris kairei</i>	CIR: Kairei; Edmond	2415–3320	Vent	Watabe & Hashimoto, 2002
<i>Rimicaris hybisae</i> sp. nov.	MCSC: Beebe; Von Damm	2300–4960	Vent	This study
<i>Shinkaicaris leurokolos</i>	OT: Minami–Ensei Knoll	~700	Vent	Kikuchi & Hashimoto, 2000; Komai & Segonzac, 2005

CIR, Central Indian Ridge; ELSC, East Lau Spreading Centre; EPR, East Pacific Rise; GoM, Gulf of Mexico; GR, Galapagos Rift; KR, Kermadec Ridge, New Zealand; LB, Lau Basin; MAR, Mid-Atlantic Ridge; MBAB, Mariana Back-Arc Basin; MCSC, Mid-Cayman Spreading Centre; OT, Okinawa Trough; SB, Sagami Bay.

venting sulphide chimneys and, at 4960 m, it is the world's deepest known vent field (Connelly *et al.*, in press). The Von Damm Vent Field (VDVF) occurs 30 km from the BVF, on the upper slopes of an off-axis massif, at 2300 m depth (Connelly *et al.*, in press). The VDVF comprises a sulphide mound (100 m diameter, 30 m height) venting predominantly clear, buoyant, high-temperature (>140°C) fluids from orifices at its peak (Connelly *et al.*, in press). Study of the fauna

inhabiting these unique vents has the potential to enhance current understanding of the dispersal, isolation, and evolution of vent taxa and patterns of vent biogeography.

The fauna at both vent fields is dominated by dense aggregations of *Rimicaris hybisae* (Figure 1), a new species of alvinocaridid shrimp, with a greatly reduced rostrum, and a four-lobed dorsal organ, similar to the photoreceptor of *Rimicaris* (Van Dover *et al.*, 1989). *Rimicaris hybisae* sp.

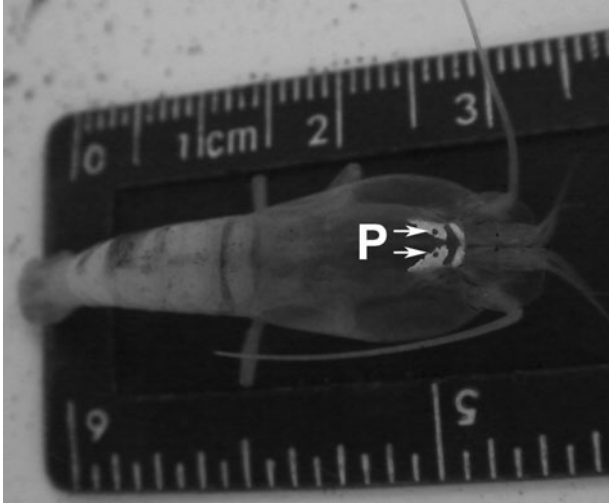


Fig. 1. *Rimicaris hybisae* sp. nov., live specimen, from the Beebe Vent Field, Mid-Cayman Spreading Centre. P denotes twin pores in the dorsal organ. 69 × 57 mm (150 × 150 DPI).

nov., described and illustrated herein, is the first taxon to be described from vent fields on the MCSC. In addition to enhancing existing knowledge about biodiversity, this record extends the known geographical range of *Rimicaris* (previously only recorded from Atlantic and Indian Ocean vents; Table 1) westwards into the Caribbean Sea, and extends the known bathymetric range of the Alvinocarididae by 872 m.

MATERIALS AND METHODS

The specimens were collected during the 44th cruise of the RRS 'James Cook' in April 2010 from Beebe (4960 m) and Von Damm (2300 m) hydrothermal vent fields on the MCSC, Caribbean. Shrimp samples were taken using a grab

(Von Damm) and suction sampler (Beebe) attached to the manoeuvrable TV grab HyBIS (Hydraulic Benthic Interactive Sampler), together with still photographs and video recordings of them *in situ* at both sites. Material for morphological study was immediately fixed in 10% neutralized formalin and subsequently transferred to 80% industrial methylated spirits on return to the laboratory. Material for molecular analysis was immediately placed in 100% ethanol.

The measurements taken for each specimen (Table 2) were measured to the nearest 0.1 mm using Vernier callipers. Post-orbital carapace length (CL) was measured from the mid-point of the posterodorsal margin to the level of the posterior margin of the orbit. Maximum total length (TL) was measured from the posterior margin of the telson to the anterior margin of the antennal scale. Maximum carapace width (CW) and maximum carapace depth (CD) were measured at the widest and deepest points of the carapace respectively. Specimen size herein is indicated by post-orbital CL.

Individuals were sexed under a dissecting microscope. Males were distinguished by an asymmetrical mesial extension on the endopod of pleopod 1 and the presence of the appendix masculina on the second pleopod (Williams, 1988). The sex of specimens CL 6.7 mm and smaller could not be determined by this method; those specimens are referred to as juveniles.

Drawings were prepared with the aid of a camera lucida mounted onto a Leica MZ8 stereomicroscope, scanned and digitally inked using Adobe® Illustrator® and a WACOM™ digitizer, as described by Coleman (2003, 2009). The material examined is deposited in the Natural History Museum, UK (NHMUK). The descriptive terminology used follows that of Komai & Segonzac (2008).

One specimen from the BVF was used for scanning electron microscopy (SEM) and was not designated as a paratype. This specimen was dehydrated through a graded ethanol series, critical point dried and sputter coated with gold palladium prior to examination with a FEI Quanta 200 scanning electron microscope at accelerating voltage of 10 kV.

Table 2. Morphological variation in *Rimicaris hybisae* sp. nov. Mid-Cayman Spreading Centre vent fields are abbreviated as VDVF (Von Damm) and BVF (Beebe).

Site	Cat. no.	Sex	Type status	CL (mm)	CW (mm)	CD (mm)	CW/CD	TL (mm)	Teeth AS ₄ left	Teeth AS ₄ right	Teeth AS ₅ left	Teeth AS ₅ right
VDVF	NHMUK 2011.8054	Female	Holotype	15.3	12.6	7.5	1.7	46.6	0	0	0	0
VDVF	NHMUK 2011.8055	Male	Paratype	9.4	8.8	5.2	1.7	32.5	0	0	1	1
BVF	NHMUK 2011.8056	Female	Paratype	10.8	9.3	5.4	1.7	39.7	0	0	0	0
BVF	NHMUK 2011.8057	Male	Paratype	10.6	8.3	7.1	1.2	35.1	1	0	4	1
BVF	NHMUK 2011.8058	Male	Paratype	10.0	7.4	6.4	1.2	36.6	0	0	0	0
BVF	NHMUK 2011.8059	Male	Paratype	10.8	8.7	6.9	1.3	37.7	0	0	1	3
BVF	NHMUK 2011.8060	Male	Paratype	10.0	8.1	5.8	1.4	33.7	0	0	1	1
BVF	NHMUK 2011.8061	Male	Paratype	11.8	9.9	7.2	1.4	36.2	0	0	0	0
BVF	NHMUK 2011.8062	Female*	Paratype	7.0	x	x	x	25.9	0	0	1	1
BVF	NHMUK 2011.8063	Male	Paratype	10.9	x	x	x	35.2	0	0	1	2
BVF	NHMUK 2011.8064	Male	Paratype	9.0	7.9	6.1	1.3	30.9	0	0	0	0
BVF	NHMUK 2011.8065	Juvenile	Paratype	6.7	4.9	4.1	1.2	22.6	0	0	2	3
BVF	NHMUK 2011.8066	Juvenile	Paratype	5.1	3.5	2.9	1.2	16.5	0	0	1	1
BVF	NHMUK 2011.8067	Juvenile	Paratype	3.9	2.7	2.2	1.2	13.1	0	0	2	2
BVF	NHMUK 2011.8068	Juvenile	Paratype	3.6	x	x	x	12.4	0	0	0	0
BVF	NHMUK 2011.8069	Juvenile	Paratype	4.6	3.4	3.3	1.0	15.1	0	0	3	1
BVF	NHMUK 2011.8070	Juvenile	Paratype	4.2	2.8	1.9	1.5	13.7	0	0	1	1

CL, post-orbital carapace length; CW, maximum carapace width; CD, maximum carapace depth; TL, total length; AS₄, abdominal somite 4, posterolateral margin; AS₅, abdominal somite 5, posterolateral margin; ovi, ovigerous; x, measurement could not be taken due to condition of carapace; *, with visibly mature ovary.

Abdominal muscle for DNA extraction was cut from the shrimp of ethanol-preserved specimens (from both the Beebe and Von Damm vent fields) and the carapace removed. Genomic DNA was extracted using the CTAB (cetyltrimethyl ammonium bromide) procedure (Doyle & Dickson, 1987). Regions of the mitochondrial genes cytochrome *c* oxidase subunit I gene (COI) and 16S ribosomal RNA, and of the nuclear 18S ribosomal RNA gene were amplified by performing polymerase chain reactions (PCR).

The COI region was amplified with universal primers LCO1490 and HCO2198 (Folmer *et al.*, 1994). The 20 µl amplification mixture contained 1X buffer reagent (200 mM Tris pH 8.8, 500 mM KCL, 0.1% Triton X-100, 2 mg/ml bovine serum albumen), 2 mM MgCl₂, 0.2 mM of each dNTP, 0.5 mM of each primer, 1 U Taq DNA polymerase (Bioline), 5 µl of template DNA and sterile H₂O to final volume. Thermal cycling conditions were: 94°C/2 minutes; followed by 5 cycles at (94°C/35 seconds; 45°C/35 seconds; 72°C/1:20 minutes) and 35 cycles at (94°C/35 seconds; 50°C/35 seconds; 72°C/1:20 minutes) with a final extension of 72°C/ 10 minutes.

For the 16S gene, PCR amplifications were performed using the universal primers 16Sar and 16Sbr (Palumbi, 1996) and a 20 µl amplification mixture: 1X reaction buffer (same as for COI), 2.5 mM MgCl₂, 0.13 mM of each dNTP, 0.38 mM of each primer, 1 U Taq DNA polymerase (Bioline), 2.5 µl of template DNA and sterile H₂O to final volume. Thermal cycling conditions were: 94°C/4 minutes; 30 cycles at (94°C/30 seconds; 52°C/1 minutes; 72°C/2 minutes) and 72°C/5 minutes.

Polymerase chain reaction amplifications of the 18S gene were performed using universal primers 18SunIF and 18SunIR (Sogin, 1990) in an amplification mixture as described for COI. Thermal cycling conditions were: 95°C/5 minutes; 30 cycles at (94°C/1 minute; 64°C/1 minute; 72°C/2:30 minutes) and 72°C/10 minutes. Negative controls were included as standard and sterile procedures were consistently followed for all PCR experiments.

Polymerase chain reaction products were purified with the ExoAP treatment by adding the following ExoAP mixture to 15 µl PCR product: 0.2 µl 10X ExoAP buffer (50 mM Bis-Tris, 1mM MgCl₂, 0.1 mM ZnSO₄), 0.05 µl 5000 U/ml Antarctic Phosphatase (New England Biolabs: Ipswich, MA), 0.05 µl 20000 U/ml Exonuclease I, and 3.7 µl sterile H₂O) and thermal-cycler incubation (37°C/60 minutes; 85°C/15 minutes). Sequencing reactions were performed using BigDye Terminator Reactions following the manufacturer's protocol (Applied Biosystems: Foster, CA) with the same primer sets used for amplifications. For COI, the thermal-cycler reaction was performed as: 94°C/30 seconds followed by 25 cycles at (94°C/15 seconds; 50°C/15 seconds; 60°C/3 minutes). For 16S and 18S the PCR conditions were identical to those described for COI, except for the use of a 52°C and 64°C annealing temperature respectively. Sequencing reaction products were purified with the AMPure magnetic bead system (Agencourt: Morrisville, NC) following the manufacturer's protocol and were subsequently run on an ABI 3730x1 DNA Analyzer (Applied Biosystems International).

The sequence strands for each gene were proof-read and assembled with CodonCode Aligner, version 3.7.1 (CodonCode Corporation, Dedham, MA, USA), to produce a continuous fragment. Sequences were compared with

those in GenBank using the nucleotide BLAST program (NCBI Basic Alignment Search Tool) and manually aligned in BioEdit (Hall, 1999). Phylogenetic trees were constructed with MEGA5 (Tamura *et al.*, 2011) using the neighbour-joining (NJ) (Saitou & Nei, 1987) and maximum-likelihood (ML) (Kimura, 1980) methods on 460- and 540-base pair (bp) alignments for COI and 18S respectively. Bootstrap values were calculated on 1000 re-sampling replicates.

GenBank accession numbers for partial sequences of the 16S, COI and 18S regions are JN850606, JN850607 and JN850608 respectively.

SYSTEMATICS

Order DECAPODA Latreille, 1802
 Infraorder CARIDEA Dana, 1852
 Superfamily BRESILOIDEA Calman, 1896
 Family ALVINOCARIDIDAE Christoffersen, 1986
 Genus *Rimicaris* Williams & Rona, 1986

Rimicaris Williams & Rona, 1986: 447 (in part); Martin & Hessler, 1990: 8; Holthuis, 1993; *Iorania* Vereshchaka, 1996: 952; Martin & Haney, 2005: 467; Komai & Segonzac, 2008: 22.

TYPE SPECIES

Rimicaris exoculata Williams & Rona, 1986

DIAGNOSIS (EMENDED)

Carapace greatly inflated laterally, distinctly broader than pleon, dorsal surface rounded, pitted with scattered, shallow punctuations. Pterygostomial expansion produced, exceeding antennal lobe, covering greater part of antennal bascicerite, rounded or blunt. Rostrum reduced to broadly rounded lobe. Eyes lacking pigment, eyestalks flattened, greatly reduced and medially fused. Antennal scale broadly oval, bearing distolateral transverse suture. Mandible with two-segmented palp, distinct separation between incisor and molar processes. Maxilla with scaphognathite greatly expanded anteriorly and conspicuously setose on dorsal and ventral surfaces. First maxilliped with greatly expanded exopod, similar to scaphognathite, conspicuously setose on dorsal and ventral surfaces. Third maxilliped with three long segments and coxa.

COMPOSITION

Rimicaris exoculata Williams & Rona, 1986 (MAR, 45°N–4°47'S), *Rimicaris kairei* Watabe & Hashimoto, 2002 (Central Indian Ridge, Kairei and Edmond vent fields) and *Rimicaris hybisae* sp. nov. (Beebe and Von Damm vent fields, MCSC, Caribbean).

Rimicaris hybisae sp. nov.
 (Figures 1–7)

TYPE MATERIAL

Holotype: adult female, CL 15.3 mm. VDVF, MCSC, Caribbean Sea; co-ordinates: 18°22.605'N81°47.875'W; water depth: 2300 m, [NHMUK 2011.8054]. Collected on the 44th voyage of RRS 'James Cook', on 18 April 2010.

Paratypes: adult male, CL 9.4 mm. VDVF, MCSC, Caribbean Sea; co-ordinates: 18°22.605'N81°47.875'W; water depth: 2300 m, [NHMUK 2011.8055]. Collected during the 44th cruise of the RRS 'James Cook', on 18 April 2010. Adult female, CL 10.8 mm; adult female with visible mature ovary, CL 7.0 mm; seven adult males, CL 7.0–11.8 mm; six juveniles, CL 3.6–6.7 mm. BVF, MCSC, Caribbean Sea;

co-ordinates: $18^{\circ}32.785'N$ $81^{\circ}43.080'W$; water depth: 4960 m, [NHMUK 2011.8056-8070]. Collected on the 44th voyage of RRS 'James Cook', on 15 April, 2010.

COMPARATIVE MATERIAL EXAMINED

Chorocaris chacei (Williams & Rona, 1986). Ten males, CL 9.7–13.0 mm [MNHN–Na17811]. MAR (Lucky Strike: Tour Eiffel; $37^{\circ}17'N$ $32^{\circ}17'W$). Collected by net, 1689 m depth.

Chorocaris vandoverae Martin & Hessler, 1990. Paratypes: seven females, CL 8.0–12.6 mm [USNM 243947]. Mariana Back Arc Basin (Alice Spring Vent Field; $18^{\circ}12.599'N$ $144^{\circ}42.231'E$). Collected by net, 3640 m depth.

Rimicaris exoculata Williams & Rona, 1986. Paratypes: ten females, CL 14.3–18.4 mm [USNM 228447]. MAR (TAG; $26^{\circ}08.18'N$ $44^{\circ}49.36'W$). Collected by dredge, 3620–2650 m depth. Paratypes: five juveniles, CL 8.4–9.2 mm [USNM 228454]. Labelled as *Rimicaris chacei*; classified as *R. exoculata* juveniles at stage B by Komai & Segonzac (2008). MAR (TAG; $26^{\circ}08.3'N$ $44^{\circ}49.6'W$). Collected by dredge, 2650–3620 m depth. Twenty-Four additional specimens: eleven males, CL 11.3–18.1; thirteen females, CL 8.8–18.4 mm from J. Copley's reference collection. MAR (TAG).

Rimicaris kairei Watabe & Hashimoto, 2002. Paratype: one female, CL 19.3 mm [USNM 1005217]. Central Indian Ridge, Rodriguez Triple Junction (Kairei Vent Field; $25^{\circ}19.16'S$ $70^{\circ}02.40'E$). Collected by suction sampler, 2454 m.

Opaepelle loihi Williams & Dobbs, 1995. Paratypes: twenty females, CL 8.0–11.7 mm [USNM 1005217]. North Pacific Ocean (Loihi Seamount; $18^{\circ}55'N$ $155^{\circ}16'W$). Collected by baited traps, 990 m depth.

DIAGNOSIS

Rostrum reduced to broadly rounded lobe, nearly reaching, reaching or slightly overreaching anterior margins of medially-fused eyes; ventral surface flat or slightly convex. Carapace and pleon with scattered minute setae; antennal lobe broadly triangular or rounded; pterygostomial lobe rounded or blunt. Fourth pleonal pleuron posterolateral margin unarmed or armed with one tooth; produced into subacute posteroventral angle. Fifth pleonal pleuron posterolateral margin unarmed marginally or armed with 1–4 teeth; produced into acute posteroventral angle. Antennae not operculiform; distolateral tooth first antennal peduncle subacute or blunt; antennal scale distolateral tooth subacute. Scaphognathite of maxilla and caridean lobe of first maxilliped bearing numerous plumose setae-like structures on dorsal and ventral surfaces; exopodal flagellum of first maxilliped completely reduced. Appendix masculina tapering distally with 7–8 spiniform setae restricted to tip. Uropodal protopod posterolateral projection triangular with acute tip.

In life, with four-lobed dorsal organ; lobes fused anteriorly; posterior lobes with paired 'pores' (Figure 1).

DESCRIPTION

Body integument smooth, firm, pitted with scattered, very shallow punctuations bearing minute scattered setae (including rostrum). Carapace (Figure 2) ovate-oblong, generally broader than deep in greatest dimensions (Table 2), distinctly broader than pleon; branchial regions distinctly inflated but to a lesser extent than that of *Rimicaris exoculata* Williams & Rona, 1986. Rostrum reduced to broadly rounded lobe; nearly reaching, reaching or slightly overreaching anterior

margins of fused eye-stalks; dorsal surface rounded, ventral surface flat or slightly convex. Carapace dorsal surface rounded; obsolescent epigastric ridge present, broader than breadth between antennal lobes, defined by conspicuous grooves extending from bases of antennal lobes; ventral margin reinforced by low submarginal ridge, most robust and farthest from margin posteriorly. Antennal lobe rounded or broadly triangular, tip blunt or subacute; anterolateral margin between antennal lobe and pterygostomial expansion slightly convex. Pterygostomial expansion produced, exceeding antennal lobe, covering greater part of antennal basicerite in larger specimens, rounded or blunt. Posterior submarginal groove present, poorly defined.

Abdomen (Figure 3A) evenly rounded dorsally, without carination; anterior three pleonal pleura unarmed marginally, posteroventral angle rounded. Fourth pleonal pleuron posterolateral margin unarmed or armed with one tooth; produced into subacute posteroventral angle. Fifth pleonal pleuron posterolateral margin unarmed marginally or armed with 1–4 teeth; produced into acute posteroventral angle. Sixth pleonal pleura 1.5–1.8 times longer than fifth in dorsal midline, 1.5 times longer than high; broadly notched for insertion of uropods; posterolateral process terminating in acute triangular tooth overlapping base of telson, posteroventral corner subacute. Armature of pleonal sternites as described for *Chorocaris chacei* Williams & Rona, 1986.

Telson (Figure 3B, C) 1.5–1.7 times length of sixth pleonal pleura in dorsal midline, slightly narrowed posteriorly, length 2.2–2.9 times greatest width; posterior margin broadly convex, bearing row of 8–35 plumose setae and 2 spines at both lateral ends; 4–8 (sometimes asymmetrical) dorsolateral spines arranged in sinuous row.

Uropods (Figure 3B) with broad rami exceeding distal margin of telson; exopod with distinct transverse suture and

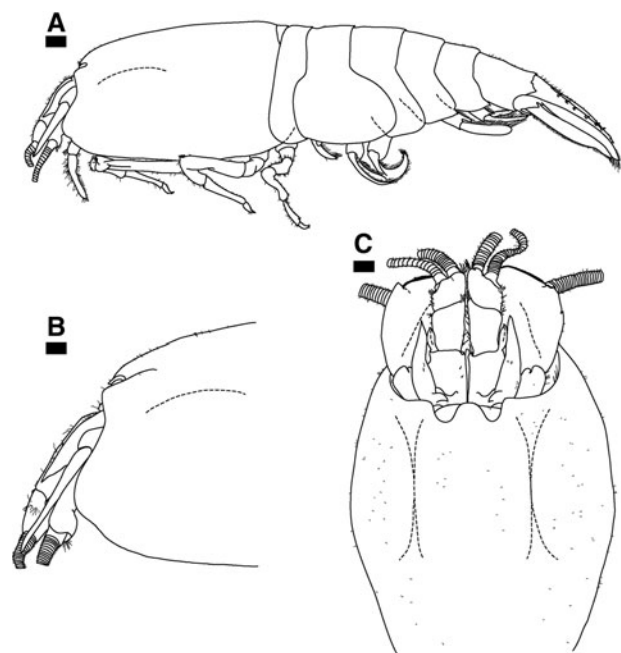


Fig. 2. *Rimicaris hybisae* sp. nov., holotype, female (carapace length 15.3 mm), [NHMUK 2011.8054], from the Von Damm Vent Field, Mid-Cayman Spreading Centre: (A) entire animal, lateral view; (B) carapace and cephalic appendages, lateral view; (C) carapace and cephalic appendages, dorsal view. Scale bars = 2 mm. 185 × 195 mm (150 × 150 DPI).

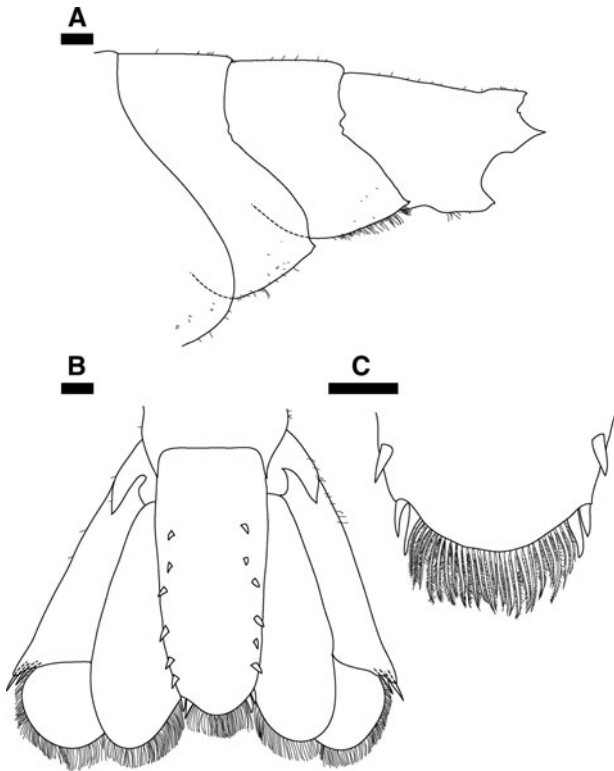


Fig. 3. *Rimicaris hybisae* sp. nov., holotype, female (carapace length 15.3 mm), [NHMUK 2011.8054], from the Von Damm Vent Field, Mid-Cayman Spreading Centre: (A) third to sixth abdominal somites, lateral view; (B) telson and uropods, dorsal view; (C) posterior margin of telson, dorsal view. Scale bars = 1 mm. 219 × 275 mm (150 × 150 DPI).

two or three small spines at distolateral angle; endopod shorter and narrower than exopod; posterolateral projection of protopod triangular with acute tip.

Eye-stalks (Figure 2C) broadly fused mesially, lacking pigmentation, anterior surface without conspicuous tubercles.

Antennae (Figure 2) of normal structure. Antennular peduncles stout, dorsoventrally depressed, mesial surface rounded, not closely approximated; first and third segments nearly equal in length, second segment slightly longer; first segment with small distomesial tooth, strong distolateral tooth subacute or blunt, reaching or overreaching midlength of second segment, dorsal face convex; stylocerite moderately slender, lateral margin slightly convex, distinctly separated from first segment, extending beyond midlength of second segment; prominent proximolateral tubercle; second segment longer than broad or nearly as long as broad with small distomesial tooth. Antennular flagella robust, thick, similar in structure, tapering, sensory setae at joints between annuli, inserted next to each other on oblique terminal margin of third article, sweeping posterolaterally in gentle arc.

Antennal peduncles (Figure 2) shorter than antennular peduncles, both exceeded by antennal scale (scaphocerite); flagella larger (all dimensions) than those of antennule and of similar structure, sensory setae at joints of annuli, sweeping in gentle curve lateral to carapace, reaching as far as midlength of fourth pleonal pleuron in holotype female. Antennal scale (Figures 2 & 4A) broadly oval; convex lateral margin ending in subacute tooth directed forward, short transverse suture extending mesially from base of tooth; plumose setae fringing broadly rounded distal margin and convex mesial margin.

Antennal basicerite bearing acute ventrolateral and ventromesial tooth.

Mouthparts (Figure 4) typical of Alvinocarididae. Mandible (Figure 4B) bearing 7 unequal acute teeth on mesial margin of broad incisor process (4 large and 3 small in holotype female), distalmost tooth distinctly separated from remaining teeth; molar process slightly upcurved, slender, unarmed, not reaching incisor; palp biarticulate, proximal article weakly curved, bearing 2 long plumose setae on distolateral margin, distal article stout, distinctly shorter than basal article, bearing numerous plumose setae of variable lengths on all margins and ventral face.

Maxillule (first maxilla) (Figure 4C) with both endites strongly curved toward mouth; coxal endite semitriangular, bearing long, dense, stiff setae; basal endite more rounded dorsally and distally, armed with row of stiff setae (shorter than those of coxal endite) and 3 rows of spines along mesial margin, each successive row becoming more regularly spaced, bearing spines in greater number and size; lateral margin bearing row of long plumose setae; palp weakly curved, slightly bilobed distally with proximal lobe bearing long plumose seta, distal lobe bearing short plumose seta.

Maxilla (second maxilla) (Figure 4D) with densely setose mesial endites; proximal endite curved, straplike lobe; distal endite composed of two lobes, separated from each other by a deep notch and suture; flanked by dorsoventrally compressed tapering palp; scaphognathite enormously expanded, numerous plumose-setae like structures conspicuous on dorsal and ventral surfaces and lateral and mesial margins, supporting dense coverage of numerous filamentous bacteria-like structures (Figure 5); posterior lobe elongate, subtriangular, convex mesial margin distinctly notched, distomesial margin fringed with very long, wiry, tangled setae, preceded by much shorter plumose setae along proximomesial margin.

First maxilliped (Figure 4E, F) with irregularly fusiform, heavily setose mesial endite; palp concealed by caridean lobe, short and bilobed, proximal lobe straplike bearing plumose setae along margins, distal lobe triangular, without ornamentation; caridean lobe broad, similar in shape and ornamentation to scaphognathite, lacking flagellum; large epipod subrectangular, curving dorsally, scattered setae on distolateral margin only.

Second maxilliped (Figure 4G) composed of six segments as in other Alvinocarididae; coxa expanded mesially, bearing numerous plumose setae on mesial margin; merus and ischium-basis fused segments moderately stout with numerous plumose setae on curved lateral surfaces, row of dorsally curved setae on nearly straight mesial surface; articulation between merus and carpus oblique; carpus short, long plumose setae on outer face; propodus with moderately long plumose setae mesially; articulation between propodus and dactylus oblique; dactylus longer than propodus, tapering to blunt distal margin, bearing very dense patch of short setae on mesial to distal margins forming brush-like structure; triangular epipod with slender rudimentary protobranch overreaching distal margin of epipod.

Third maxilliped (Figure 4H) overreaching antennal peduncle by one-third of ultimate segment, comprising coxa and three long segments; coxa heavily setose, with large, bilobed epipod; epipod without strap-like process; antepenultimate segment longest (consisting of fused basis–ischium–merus, but fusion between basis and ischium incomplete with partial suture on dorsal surface and corresponding

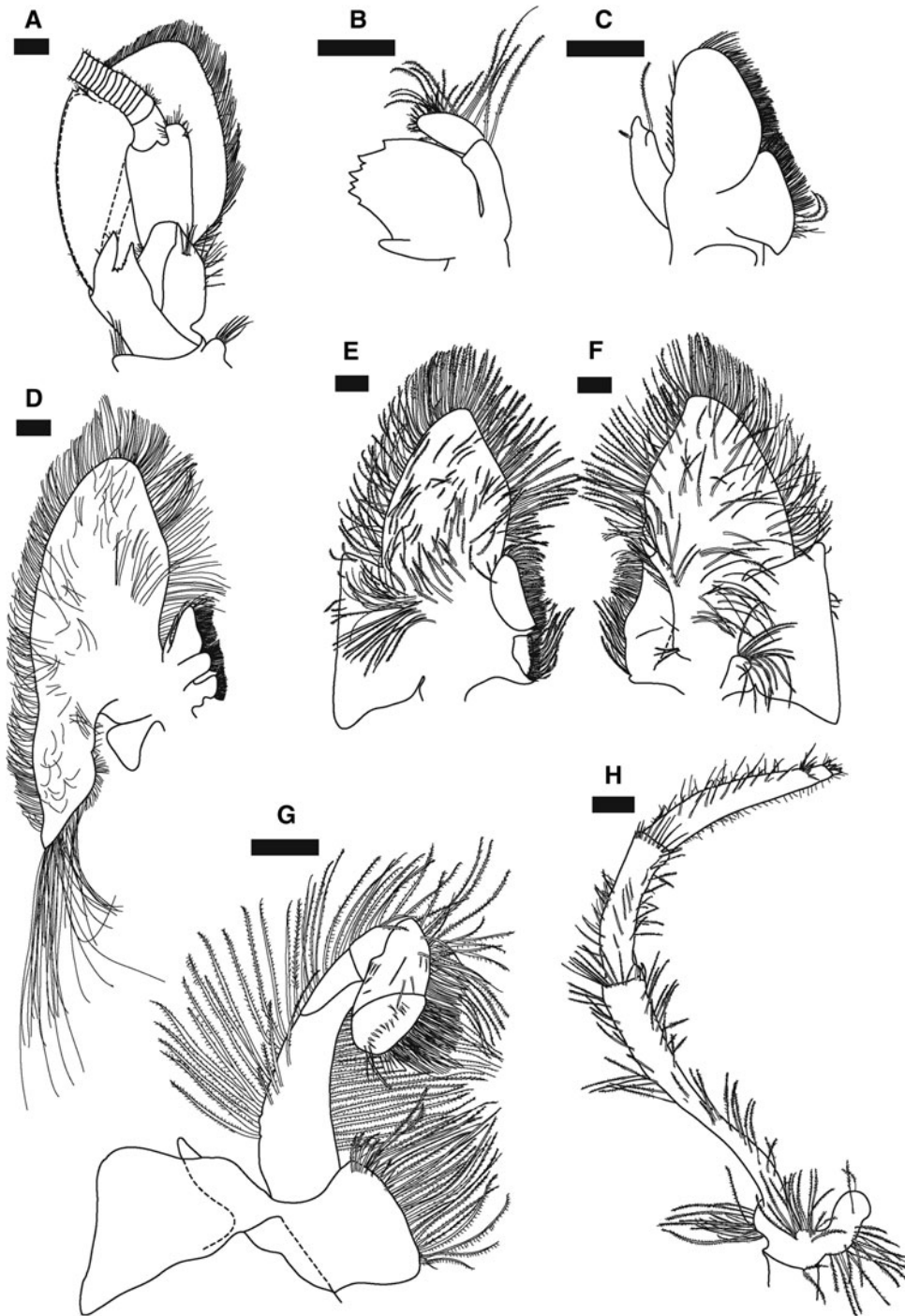


Fig. 4. *Rimicaris hybisae* sp. nov., holotype, female (carapace length 15.3 mm), [NHMUK 2011.8054], from the Von Damm Vent Field, Mid-Cayman Spreading Centre: (A) right antennal peduncle, scale and proximal part of flagellum, ventral view; (B) right mandible, dorsal view; (C) right maxillule (first maxilla), ventral view; (D) right maxillule (second maxilla), ventral view; (E) right first maxilliped, ventral view; (F) right first maxilliped, dorsal view; (G) right second maxilliped, ventral view; (H) right third maxilliped, lateral view. Scale bars = 1 mm. 255 × 373 mm (150 × 150 DPI).

indentations on lateral and mesial margins), dorsoventrally flattened, sinuously curved in dorsal view, armed with 1–2 spines at distolateral ventral angle, bearing numerous plumose setae on margins, dense cluster of long setulose setae on low elevation at proximomesial section, elevation length one-fifth of segment length, mesial margin notched where elevation terminates; penultimate segment with plumose setae on dorsolateral surfaces, rows of very dense, short, stiff setae on ventromesial face distal to one-third

length; ultimate segment strongly curved, stout, about 1.5 times longer than penultimate segment, tapering distally to truncate tip bearing several terminal spines (7 on holotype female), trigonal in cross-section, lateral surface longitudinally carinate, with row of stiff setae, ventromesial surface flat with rows of very dense, short, stiff setae.

Branchial formula identical to that described in *Rimicaris* and *Chorocaris* (Williams & Rona, 1986; Williams, 1988; Martin & Hessler, 1990; Komai & Segonzac, 2008).

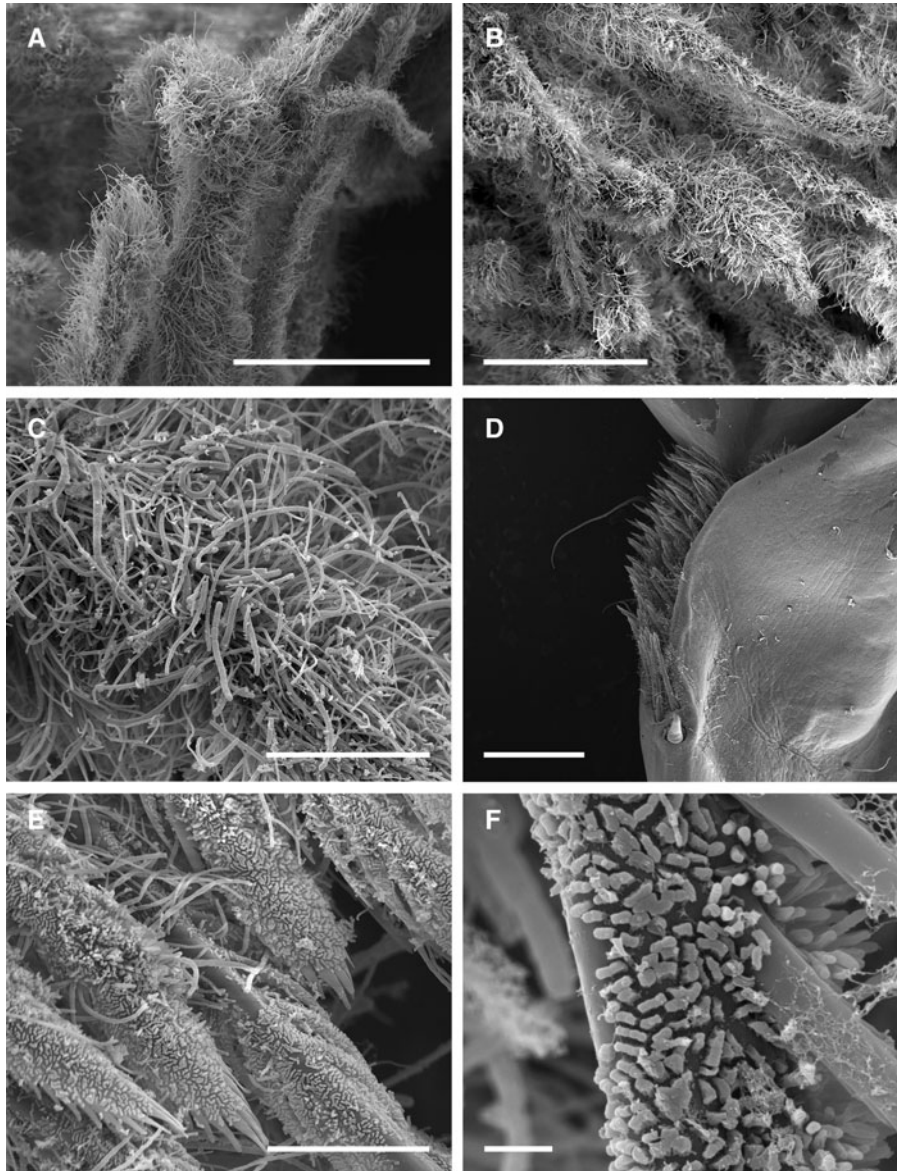


Fig. 5. *Rimicaris hybisae* sp. nov., male, from the Beebe Vent Field, Mid-Cayman Spreading Centre. Scanning electron micrographs of: (A) plumose-setae like structures, margin of maxilla; (B) plumose-setae like structures, ventral surface of maxilla; (C) close-up of filamentous bacteria-like structures, maxilla; (D) carpal brush on distal segment of carpus, first pereopod; (E) close-up of carpal brush setae and bacteria-like structures; (F) close-up of dorsal surface of carpal brush seta and setules with filamentous and non-filamentous bacteria-like structures. Scale bars: A = 300 μm ; B, D = 200 μm ; C = 50 μm ; E = 20 μm ; F = 2 μm . 227 \times 296 mm (150 \times 150 DPI).

Pereopods (Figure 6) without epipods. First pereopod typical of family, polymorphic, smaller than succeeding pereopods, typically folded upon itself at mero-carpal joint, reaching tip of antennal peduncle when extended; carpus distal margin obliquely truncate for accommodation of proximal part of palm, mesial face bearing grooming apparatus (carpal brush *sensu* Martin *et al.*, 1998: figures 1–3) comprised a triangular patch of serrate setae (some setules also serrate) arising from a recessed area, a row of distally-orientated serrate setae just proximal to the setal patch, and a stout spine proximal to the row of setae (Figure 5D); carpal brush setae bearing numerous filamentous and non-filamentous bacteria-like structures (Figure 5E, F); dactyl slightly overreaching fixed finger.

Second pereopod more slender than first, slightly twisted, just overreaching tip of antennal peduncle if extended;

merus and ischium unarmed; carpus and chela much weaker than those of first pereopod.

Third to fifth pereopods similar in structure, moderately stout, normally folded at mero-carpal articulation, meri and carpi somewhat twisted, meri and ischia unarmed, decreasing in length from third to fifth (e.g. 20.5–18.0–17.1 mm respectively in holotype female); carpi 1.1–1.2 length of propodi in third pereopod, 0.9–1.1 in second, 0.7–0.9 in third; propodi increasing in length from third to fifth, with irregular double (more or less) row of spinules on ventral surface and normally 4 spinules on ventrodorsal margin; dactyli subconical, 0.2 length of propodi, each terminating in robust, curved, corneous unguis, flexor surface bearing marginal and plantar rows of distally curved, corneous spinules, increasing in strength distally.

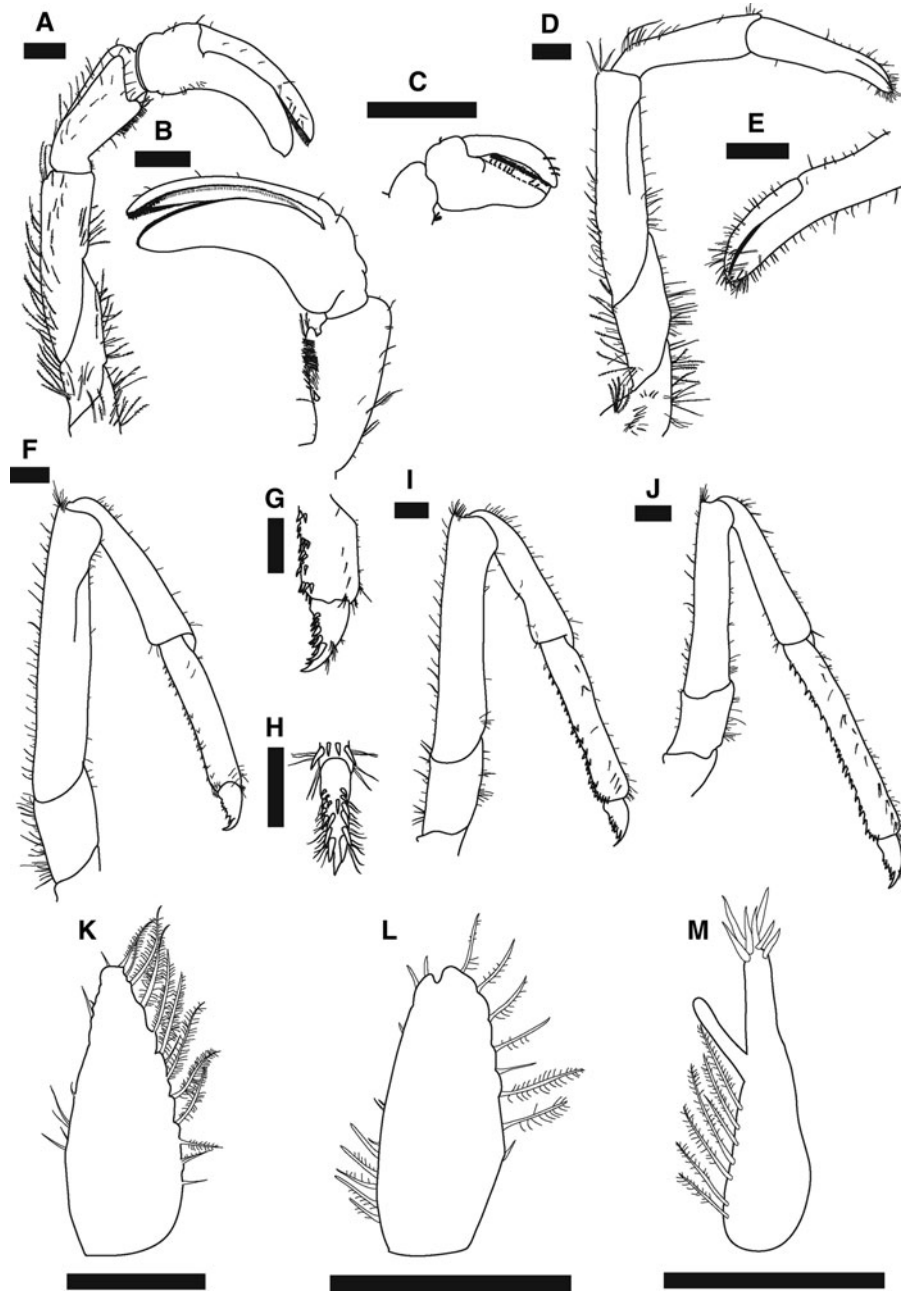


Fig. 6. *Rimicaris hybisae* sp. nov.: (A, B & D–K) holotype, female (carapace length (CL) 15.3 mm), [NHMUK2011.8054], from the Beebe Vent Field, Mid-Cayman Spreading Centre (MCSC); (C) paratype, male (CL 5.1 mm), [NHMUK2011.808066], from the Von Damm Vent Field (VDVF), MCSC; (L–M) paratype, male (CL 9.4 mm), [NHMUK2011.8055], from VDVF, MCSC; (A) right first pereopod, lateral view; (B) chela and carpus of right first pereopod, mesial view; (C) chela of left first pereopod, inner view; (D) right second pereopod, lateral view; (E) chela of right second pereopod, mesial view; (F) right third pereopod, lateral view; (G) dactylus and distal part of propodus of right third pereopod, lateral view; (H) dactylus of third right pereopod, posterior view; (I) right fourth pereopod; (J) right fifth pereopod; (K) endopod of right first pleopod, female, ventral view; (L) endopod of left first pleopod, male, ventral view; (M) appendix masculina and appendix interna, lateral view. Scale bars = 1 mm. 250 × 359 mm (150 × 150 DPI).

First pleopod bearing sexually dimorphic endopod; in female, endopod (Figure 6K) simple, terminating in blunt apex; in male, endopod (Figure 6L) with asymmetrical distal notch separating much produced mesial lobe from smaller distolateral lobe. Second pleopod with slender appendix interna without cincinnuli; appendix masculina (Figure 6M) (males) tapering distally, bearing 7–8 spiniform setae distally. Third and fourth pleopods each with slender appendix interna, third without cincinnuli, fourth bearing a few tiny cincinnuli at tip. Fifth pereopod with stout, more robust

appendix interna with many cincinnuli in subapical mesial cluster.

COLORATION

In life, carapace mostly pale translucent white; internal tissues neutral to greyish; integument of juveniles more translucent, oily orange globules visible underneath, tissues more orange.

Specimens from the BVF with rust-coloured deposits, most notably under antero-lateral area of carapace, on ventral

surfaces of thorax, abdomen, frontal region, all areas of maxilla and first maxilliped; blackening common under antero-dorsal area of carapace (Figure 1), on tips of dactyli, third maxilliped and carpal brush; black sparkly particles clustered between mouthparts. Specimens from the VDVF are all 'clean' in appearance, lacking the rust-coloured deposits, blackened areas and black particulate matter observed on specimens collected from the BVF.

Eyes lacking coloration and pigment; juveniles brown pigmentation present in proximal area of fused eyes; orange tint in smallest juveniles (CL 3.6–4.2 mm).

Four-lobed dorsal organ (Figure 1) highly reflective with a slight pink tint; reflective property and coloration not preserved in death or after chemical fixation; white pigmentation visible in specimens preserved in 100% ethanol; brown pigmentation under carapace in juveniles (where dorsal organ is located in adults).

Gills typically bright white.

Third to fifth pereopods each ending in pale brown dactylus visible in preservation.

VARIATION

Specimens from both vent fields exhibit similar morphology. Variation of some features within and between specimens, most notably the posteroventral angle and armature of fourth and fifth pleonal pleura (Table 2), the number of dorsolateral spines on the telson and distolateral spines on the exopod of the uropod (noted above). Slight intra- and inter-specimen variation also observed in the degree of extension of the lateral lobe of the endopod of the first pleopod in males and the shape and reach of the distolateral tooth on the first antennal peduncle (noted above). Variability in these features does not appear to occur in any consistent combination, or in relation to sex or size.

The chela of the first pereopod appears to be polymorphic. In 16 specimens the chela of the first pereopod is slightly recurved, slender and somewhat delicate (Figure 6B). In contrast, one male (CL 5.1 mm), [NHMUK 2011.8066], BVF, has a stouter, more robust chela (Figure 6C).

No variation was observed between the female with a visible mature ovary (CL 7.0 mm) [NHMUK2011.8062] and other females.

One male specimen (CL 9.4 mm), [NHMUK 2011.8055] (Figure 7), from the VDVF has an acutely pointed rostrum, antennal lobe and pterygostomial expansion. All other specimens have a rounded or blunt antennal lobe and pterygostomial expansion with a rounded rostrum.

In juveniles (Table 2), carapace less inflated; rostrum distinctly pointed downwards, lateral and dorsal surfaces convex, anterior margin ornamented with small, distally-projecting setae; smallest juveniles (CL 3.6–4.2 mm) [NHMUK 2011.8067, NHMUK 2011.8068, NHMUK 2011.8070] medially-fused eyes with tiny tubercle on anterior surface medially.

Variation in coloration noted above.

COMPARATIVE REMARKS

The present new species has been mentioned previously in the literature as 'a new morphospecies of alvinocaridid shrimp' and 'the MCSC vent shrimp' (Connelly *et al.*, in press) (Figures 4 & 5A). It is morphologically most similar to *Rimicaris exoculata*, *R. kairei* and *Chorocaris chacei* because of the reduction of the rostrum to a broadly

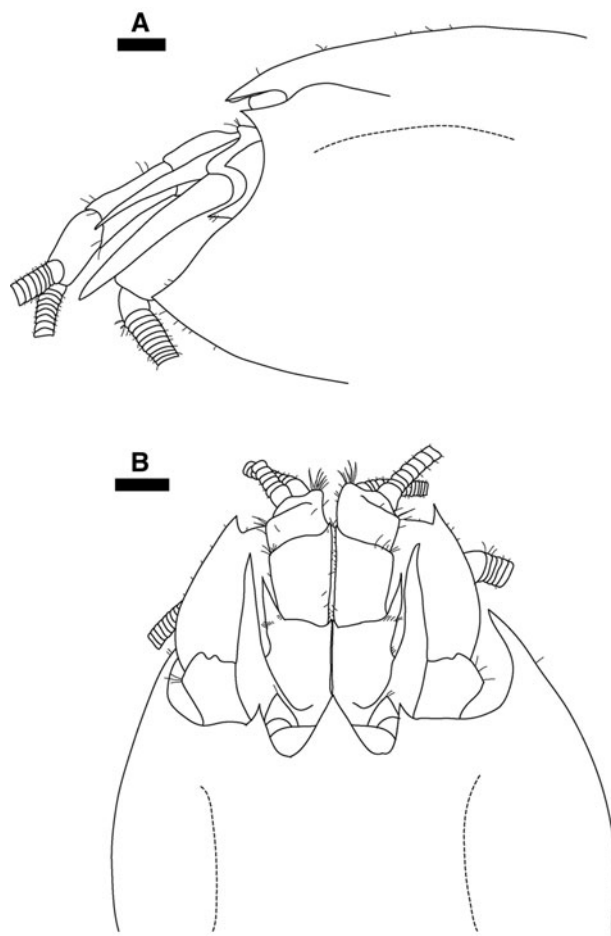


Fig. 7. *Rimicaris hybisae* sp. nov., paratype, male (carapace length 9.4 mm), [NHMUK 2011.8055], from the Von Damm Vent Field, Mid-Cayman Spreading Centre: (A) Carapace and cephalic appendages, lateral view; (B) carapace and cephalic appendages, dorsal view. Scale bars = 1 mm. 129 × 197 mm (150 × 150 DPI).

rounded lobe and nonacuminate antennal lobe of the carapace, the presence of plumose-seta like structures on the dorsal and ventral surfaces of the scaphognathite of the maxilla and caridean lobe of the first maxilliped, and an appendix masculina armed with distal setae only. The new species is easily differentiated from *C. chacei* by the more inflated anterolateral region of the carapace, possession of a four-lobed dorsal organ, and acute tip of the uropodal proto-pod. A four-lobed dorsal organ and inflated carapace are known only for *R. hybisae* sp. nov. and the two other known *Rimicaris* species. The new species, however, is the first example within the genus to possess paired 'pores' on the posterior lobes of the dorsal organ. Furthermore, the carapaces of the new species and *R. exoculata* are ornamented with setae, whereas there are no setae on the carapace of *R. kairei*. The carapace of *R. hybisae* sp. nov. is slightly less inflated than that of *R. exoculata* and *R. kairei*. Of the three known *Rimicaris* species, the carapace is most strongly inflated in *R. kairei*. Other differentiating characters between *R. hybisae* sp. nov. and the two other *Rimicaris* species include the structure of the antennae and extent to which the eyes are fused and rostrum reduced. The characters differentiating between the new species and its closest relatives are discussed below.

DISTRIBUTION AND HABITAT

Known only from the type locality, the Von Damm (2300 m) and Beebe (4960 m) hydrothermal vent fields, Mid-Cayman Spreading Centre, Caribbean Sea. For preliminary descriptions of both vent fields, Connelly *et al.* (in press). Observed at the BVF in dense aggregations on the vent chimneys (>2000 individuals m⁻², Connelly *et al.*, in press), and with high abundances of anemones around crevices issuing visible diffuse flow in the central area of the sulphide mound (Connelly *et al.*, in press). At the VDVF, in dense aggregations around actively venting orifices of the edifice peak (>2000 individuals m⁻², Connelly *et al.*, in press), along with another, numerically subordinate, morphotype of shrimp.

ETYMOLOGY

From the name of the British manoeuvrable TV grab HyBIS, in celebration of her first, and highly successful, scientific mission. The gender is feminine.

MOLECULAR PHYLOGENY

Partial sequences of the mitochondrial COI (460 bp) and 16S (549 bp) regions and the nuclear 18S (576 bp) region of *Rimicaris hybisae* sp. nov. were consistent among specimens from both sites. Unique and fixed mutations were observed in the partial sequences of the COI and 18S regions. Based on NJ and ML phylogenetic analyses for COI sequences available in GenBank, *R. hybisae* sp. nov. exhibits the smallest evolutionary distance to *Alvinocaris* sp. [AY163260.1], with a 1.96% divergence from this species, compared with a 7.39% divergence between *R. hybisae* sp. nov. and both *R. exoculata* and *Chorocaris chacei*. In the 16S region, no substitution was found between *R. hybisae* sp. nov. and *C. chacei* (0% divergence) based on the 300-bp sequence in GenBank for the latter species, whereas *R. hybisae* sp. nov. exhibits 0.47% divergence (424 bp) with *R. exoculata*. Phylogenetic analyses on the 300-bp region common among the species shows high bootstrap values (100% and 95% for NJ and ML methods respectively) separating the *R. exoculata*–*R. hybisae* sp. nov.–*C. chacei* clade from the other alvinocaridid species. Within this clade *R. hybisae* sp. nov. and *C. chacei* are separated from *R. exoculata* (88% bootstrap support, NJ and ML methods).

In GenBank, partial sequences of the 18S region for alvinocaridids are limited to *Rimicaris exoculata*, *Chorocaris chacei*, *Alvinocaris muricola* Williams, 1988 and *Alvinocaris*

longirostris Kikuchi & Ohta, 1995. Because there is only a single partial sequence in the database for both *R. exoculata* and *C. chacei*, it is not known if the mutations observed in the 18S partial sequences of these species are fixed within each of these species. Based on a 540-bp alignment, NJ and ML phylogenetic trees place *R. hybisae* sp. nov. in the same clade as *R. exoculata* and *C. chacei* (88% and 90% for NJ and ML methods respectively) (Figure 8). For the available 18S partial sequences the new species is closest in evolutionary distance to *C. chacei* (0.56% and 0.88% divergence with *C. chacei* and *R. exoculata* respectively).

DISCUSSION

The presence of unique and fixed mutations in the partial sequences of the COI and 18S region suggest that *Rimicaris hybisae* sp. nov. is genetically distinct from all other species in the GenBank database. This supports the morphological evidence that *R. hybisae* is a new species. Although morphological variation was present amongst the specimens studied, partial sequences of the COI, 16S and 18S regions were consistent between specimens, confirming that those analysed from both MCSC vent fields are monospecific.

Based on morphology, and supported by the results from the molecular analyses, *Rimicaris hybisae* sp. nov. belongs within the *Rimicaris*–*Chorocaris*–*Opaepele* clade. Common features within this complex include a greatly reduced rostrum, broadly fused eyes, three or greater rows of accessory spines on the ventral surfaces of the dactyli of the third to fifth pereopods, the unarmed ischium of the third to fifth pereopods, and sinuous rows of dorsolateral spines on the telson (Komai & Segonzac, 2005, 2008). *Rimicaris hybisae* sp. nov. differs from *Opaepele* in possessing a more reduced rostrum, inflated carapace and non-acuminate antennal and pterygostomial lobes (Lunina & Vereshchaka, 2010). The assignment of the new species to a genus was a complex issue. Many morphological characteristics of *Chorocaris* are shared with *Rimicaris* (see Martin & Hessler, 1990; Komai & Segonzac, 2008), as exemplified by the original assignment of *C. chacei* to the genus *Rimicaris* by Williams & Rona (1986). However, a suite of morphological traits distinguish the two genera.

Two of the most striking morphological features of *Rimicaris hybisae* sp. nov. are its inflated carapace and four-lobed dorsal organ. These features are, to date, unique to

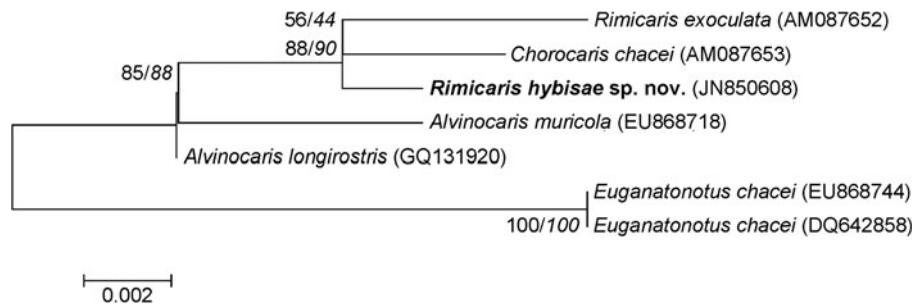


Fig. 8. Neighbour-joining tree of the Alvinocarididae based on a 540-base pair alignment of partial nucleotide sequences from the nuclear 18S ribosomal RNA region with *Eugeanatonotus chacei* Chan & Yu, 1991 (Nematocarcinidae) as an outgroup. Evolutionary distances computed using the Jukes–Cantor method (Jukes & Cantor, 1969) are represented by branch length; scale bar is proportional to inferred nucleotide divergence. Bootstrap support calculated on 100 re-sampling replicates is shown by the numbers along the branches (neighbour joining, roman text; maximum-likelihood, italic text). GenBank accession numbers are given after species names.

Rimicaris and support placement of the new species within the genus. The carapace is marginally less inflated than that of *R. exoculata* (which is less inflated than that of *R. kairei* Watabe & Hashimoto, 2002 (authors, personal observation)). *Rimicaris hybisae* sp. nov. can be further distinguished from *R. exoculata* and *R. kairei* by the presence of paired 'pores' on the posterior lobes of the dorsal organ. Variability has been documented in the shape of the dorsal organ in adult specimens of *R. exoculata* (O'Neill *et al.*, 1995: Figure 5), but the presence of 'pores' is, to our knowledge, a feature unique to *R. hybisae* sp. nov. The 'dorsal eye' of *R. exoculata* (and presumably *R. kairei*) is an extremely efficient photoreceptor, used for detecting light emitted from the vents (Pelli & Chamberlain, 1989; Van Dover *et al.*, 1994, 1996; O'Neill *et al.*, 1995). Shiny anterior spot-like organs have been described inside the carapaces of *Chorocaris chacei*, *Mirocaris fortunata* Martin & Christiansen, 1995, *Alvinocaridinides formosa* Komai & Chan, 2010, and species of *Opaepele* and *Nautilocaris* Komai & Segonzac, 2004 (Desbruyères *et al.*, 2006; Tsuchida *et al.*, 2008; Komai & Chan, 2010). These spot-like organs may be homologous to the 'dorsal eye' found in species of *Rimicaris* (Kuenzler *et al.*, 1997; Lakin *et al.*, 1997; Komai & Chan, 2010), but are smaller and do not comprise four lobes. Histological examination of the dorsal organ, requiring the collection of further specimens, would be necessary to determine if a dorsal photoreceptor has coevolved in *R. hybisae* sp. nov.

Martin & Hessler (1990) hypothesized that *Rimicaris*, with its inflated carapace, opercular frontal region, dorsal organ, and dramatically reduced rostrum, is a derived genus that stemmed from *Chorocaris* or another morphologically similar deep-sea shrimp. The presence of an inflated carapace and four-lobed dorsal organ in *R. hybisae* sp. nov. suggest that this species may be more derived than species of *Chorocaris*. In contrast, the armature of the fourth and fifth pleonal pleura is a conservative feature, also found in more conservative genera (*Opaepele*, *Alvinocaridis* and *Shinkaicaris*: Komai & Segonzac, 2005; Komai *et al.*, 2007). Within *Rimicaris* and *Chorocaris*, the fourth and fifth pleonal pleura are subacutely or acutely pointed only at their posteroventral angles and are not marginally armed (Komai & Segonzac, 2008).

Based on analyses of the COI gene (600 bp), Shank *et al.* (1999) proposed that *Chorocaris* is a paraphyletic assemblage with *C. chacei* being more closely related to *Rimicaris exoculata*. This is supported by the most recent and complete molecular phylogeny of the Alvinocarididae based on COI (600 bp), whereby *C. chacei* clusters with *R. exoculata* with strong statistical support (100% bootstrap values: Zelnio & Hourdez, 2009). In *R. hybisae* sp. nov., *Rimicaris* species and *C. chacei*, the scaphognathite of the maxilla and caridean lobe of the first maxilliped are ornamented with numerous plumose-seta like structures on both their dorsal and ventral surfaces, whilst in *C. vandoverae* Martin & Hessler, 1990 and *C. paulexa* Martin & Shank, 2005, their ventral surfaces are nearly naked (Martin & Hessler, 1990; Martin & Shank, 2005; Komai & Segonzac, 2008). The appendix masculina of *R. hybisae* sp. nov., *C. chacei* and the *Rimicaris* species bear distal setae only, whereas setae extend onto the dorsal surface of the appendix masculina in *C. vandoverae*. Komai & Segonzac (2008) suggest that this feature supports the close proximity between *C. chacei* and *Rimicaris* species and also consider *C. chacei* as a possible sister species of the

Rimicaris clade. *Rimicaris hybisae* sp. nov., with shared morphological affinities with both *C. chacei* and *Rimicaris*, further supports the proximity between these taxa, although the more conservative features of the new species are not shared with either genus.

Features common to *Chorocaris* and absent in *Rimicaris exoculata* and *R. kairei* include: possession of a strong distolateral tooth, a small distomesial tooth, and a prominent proximalateral tubercle on the first segment of the antennal peduncle; a clear separation of the stylocerite from the antennal peduncle; a carpal brush on the first pereopod; the armed antepenultimate segment of the third maxilliped; blunt or subacute antennal teeth; an increase in length from the third to fifth pereopods and a well-developed, functional appendix interna bearing cincinnuli on the fifth pleopod only (Martin & Hessler, 1990; Komai & Segonzac, 2008; authors, personal observation). These traits are also exhibited by *R. hybisae* sp. nov., revealing some morphological affinities with the genus *Chorocaris*.

Notable autapomorphies of *Rimicaris* are: the complete fusion of the eyes into a transverse ocular plate; the reduction of the rostrum to a broadly rounded lobe, fitting closely to the posterior concavity of the ocular plate; the formation of the antennal components into an operculum-like structure and the presence of a mat of dense spinules on the flexor surface of the propodi of the third to fifth pereopods (Martin & Hessler, 1990; Komai & Segonzac, 2008). These features were not observed in *R. hybisae* sp. nov., despite the examination of specimens comparable in size to adult *R. exoculata* and *R. kairei*.

In consideration of the morphological and molecular evidence presented here, it appears that *Rimicaris hybisae* sp. nov. may be intermediate between *Chorocaris chacei* and *R. exoculata*. Given the uncertainty concerning the *Chorocaris* genus, the establishment of a new genus for *R. hybisae* sp. nov. is premature. In reverence to the fact that *C. chacei* was originally *Rimicaris chacei*, the assignment of *R. hybisae* sp. nov. to the genus *Rimicaris* is the most conservative approach available. As Komai & Segonzac (2008) have already suggested, a comprehensive and extensive molecular phylogenetic analysis of the whole *Rimicaris–Chorocaris–Opaepele* clade is required to clarify the relationships between these taxa. Such a study has the potential to improve markedly our current understanding of the evolution, radiation and biogeographical patterns of these shrimp among deep-sea chemosynthetic environments in the world's oceans.

Morphological variation is well documented in the Alvinocarididae and is acknowledged as common within alvinocaridid species (e.g. Kikuchi & Ohta, 1995; Martin & Shank, 2005; Komai & Segonzac, 2008). The variation described between specimens of *Rimicaris hybisae* sp. nov. does not appear to be related to size, gender, or collection site. Polymorphism of the chela of the first pereopod has also been described for species of *Chorocaris* (Martin & Shank, 2005; Komai & Segonzac, 2008). Specimens of *C. chacei* from Lucky Strike (one female in Komai & Segonzac, 2006: Plate 3, p. 241; one male in Komai & Segonzac, 2008: figure 9F, G) have been shown with an acutely pointed rostrum, antennal lobe and pterygostomial expansion; these specimens were considered by Komai & Segonzac (2008) to be an aberrant form of the species, and although this identification may not be fully justified, it is reasonable given the sampling location. Consequently the specimen illustrated in Figure 7

[NHMUK 2011.8055] is interpreted here as an aberrant form of *R. hybisae* sp. nov. The 'clean' appearance of specimens from the VDVF and presence of black and rust-coloured deposits on specimens from the BVF may be due to the differences in the chemistry of their respective habitats (Connelly *et al.*, in press).

Results from analysis of 460 bp of the COI region for *Rimicaris hybisae* sp. nov. suggest that for this gene, the new species is genetically closest to *Alvinocaris* sp. [AY163260.1] (*Alvinocaris methanophila* Komai, Shank & Van Dover, 2005 from Blake Ridge; see Van Dover *et al.*, 2003). This result does not fit well with the evidence from morphology, 16S and 18S, thus highlighting the potential problem of relying on COI for genetic barcoding. While it is possible that there is a problem with the COI sequence for *Alvinocaris* sp., a second possibility is that there is more than one gene in the COI region. Work is in progress to test the second hypothesis (Plouviez *et al.*, unpublished data). In the 16S region, *R. hybisae* sp. nov. exhibits fixed mutations that are identical to those observed in *Chorocaris chacei*. However, it was not possible to establish if these species share an identical nucleotide sequence for more than 300-bp of the 16S region. Results from analysis of the 18S region suggest that, for this slow-evolving gene, *R. hybisae* sp. nov. is closest in evolutionary distance to *C. chacei*, although this is based on a 576-bp alignment only.

Rimicaris hybisae sp. nov. is easily differentiated from *Chorocaris chacei* by the more inflated anterolateral region of the carapace, possession of a four-lobed dorsal organ, and acute tip of the uropodal protopod. Like juveniles of *C. chacei*, juveniles of *R. hybisae* sp. nov. bear plumose setae on the posterior margin of the telson, a conspicuous distolateral tooth on the first segment of the antennal peduncle and a carpal brush on the first pereopod. In a recent review, Komai & Segonzac (2008) noted differences in the morphology of ovigerous females in species of *Chorocaris* and *Rimicaris*. Samples of ovigerous females of *R. hybisae* sp. nov. are required to enable their morphology to be described and compared to other species.

In summary, morphological features and molecular analyses indicate that *Rimicaris hybisae* sp. nov. belongs within the *Rimicaris*-*Chorocaris*-*Opaepele* clade, closest to and intermediate between *C. chacei* and *R. exoculata*. The inflated carapace and four-lobed dorsal organ of *R. hybisae* sp. nov. are diagnostic and distinguishing morphological features, previously recorded only in *R. exoculata* and *R. kairei*, which support placement of the new species within the genus *Rimicaris*. The molecular evidence reported here confirms that *R. hybisae* sp. nov. is a genetically distinct species, closest to *C. chacei*, which was originally placed in the genus *Rimicaris* as *R. chacei* (Williams & Rona, 1986). Phylogenies of the Alvinocarididae suggest that *Chorocaris* is polyphyletic, with *C. chacei* more closely related to *Rimicaris* than to other *Chorocaris* species (Shank *et al.*, 1999; Zelnio & Hourdez, 2009). Moreover, on the basis of morphological traits shared between the two taxa, it has been proposed that *C. chacei* is a sister species of *Rimicaris* (Komai & Segonzac, 2008). The most conservative approach is therefore to expand the diagnosis of *Rimicaris* to incorporate *R. hybisae* sp. nov., rather than to erect a new genus in the absence of highly corroborated phylogeny. The presence of features previously considered diagnostic of *Rimicaris* spp. and *C. chacei* in *R. hybisae* sp. nov., and the low genetic divergence between these taxa

suggest that reassimilation of *C. chacei* within *Rimicaris* could also be considered in the future.

BIOGEOGRAPHY

Rimicaris hybisae sp. nov. represents the third named species in the genus, all of which are known only from hydrothermal vents in a particular area (Table 1).

Rimicaris kairei is so far restricted to its type locality in the Indian Ocean, whereas *R. exoculata* is present at vents along the MAR, where it occurs sympatrically with *Chorocaris chacei* (Table 1). *Rimicaris hybisae* sp. nov. extends the distribution of the genus approximately 4000 km westwards into the Caribbean and increases the bathymetric range of the Alvinocarididae by 872 m to 4960 m. This may also be the first record of the Alvinocarididae in the Caribbean. Escobar-Briones & Villalobos Hiriart (2003) reported an indeterminate species of *Alvinocaris* from non-chemosynthetic environments on the Banco Chinchorro, northern Caribbean, at depths of 176–203 m but provided no molecular evidence or catalogue details.

Before the complete closure of the Isthmus of Panama by 3.1 Ma (Burton *et al.*, 1997), a deep-water connection existed between the eastern Pacific and Caribbean. Martin & Hessler (1990) proposed that the presence of *Chorocaris* species in the Atlantic and Pacific indicates a faunal connection between the eastern Pacific and Mid-Atlantic vents. However, no specimens of *Rimicaris* have been collected from the Pacific Ocean, despite numerous active surveys at Pacific vents. In addition, *Chorocaris* may be paraphyletic, with the Atlantic species (*C. chacei*) belonging to *Rimicaris*. The recent discovery of hydrothermal vents and chemosynthetic communities on the MCSC provides an opportunity to test Martin & Hessler's (1990) hypothesis. The presence of shrimp-dominated communities at the BVF and the VDVF also indicate that the MCSC vent fauna shares similarities with MAR vent fauna. The descriptions of other novel taxa at MCSC vents (Nye *et al.*, unpublished data) will further elucidate the factors determining vent biogeography of this region.

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REFERENCES

- Ahyong S.T. (2009) New species and new records of hydrothermal vent shrimps from New Zealand (Caridea: Alvinocarididae, Hippolytidae). *Crustaceana* 82, 775–794.
- Ballard R.D., Bryan W., Dick H., Emery K.O., Thompson G., Uchupi E., Davis K.E., De Boer J., Delong S.E., Fox P.J., Spydell R., Stroup J., Melson W.G. and Wright R. (1979) Geological and geophysical investigation of the Mid-Cayman Rise Spreading Centre: initial results and observations. *American Geophysical Union, Maurice Ewing Series* 2, 65–95.
- Beltenev V., Ivanov V., Rozhdestvenskaya I., Cherkashev G., Stepanov T., Shilov V., Davydov M., Laiba A., Kaylio V., Narkevsky E., Pertsev A., Dobretzova I., Gustaytis A., Popva Y., Amplieva Y., Evrard C., Moskalev L. and Gebruk A. (2009) New data about hydrothermal fields on the Mid-Atlantic Ridge between 11°–14°N: 32nd Cruise of R/V Professor Logatchev. *InterRidge News* 18, 13–17.
- Burton K.W., Ling H. and O'Nions R.K. (1997) Closure of the Central American Isthmus and its effect on deep-water formation in the North Atlantic. *Nature* 383, 382–385.
- Calman W.T. (1896) On deep-sea Crustacea from the south-west of Ireland. *Transactions of the Royal Irish Academy* 31, 1–20.
- Chan T.-Y. and Yu H.-P. (1991) *Eugonatonotus chacei* sp. nov., second species of the genus (Crustacea, Decapoda, Eugonatonotidae). *Bulletin du Muséum National d'Histoire Naturelle* 13, 143–152.
- Christoffersen M.L. (1986) Phylogenetic relationships between Oplophoridae, Atyidae, Pasiphaeidae, Alvinocarididae fam. n., Bresiliidae, Psalidopodidae and Disciidae (Crustacea Caridea Atyoidea). *Boletim de Zoologia, Universidade de São Paulo* 10, 273–281.
- Christoffersen M.L. (1990) A new superfamily classification of the Caridea (Crustacea: Pleocymata) based on phylogenetic pattern. *Zeitschrift für Zoologische Systematik und Evolutionsforschung* 28, 94–106.
- Coleman C.O. (2003) 'Digital inking': how to make perfect line drawings on computers. *Organisms Diversity & Evolution* 3 Electronic Supplement 14, 1–14.
- Coleman C.O. (2009) Drawing setae the digital way. *Zoosystematics and Evolution* 85, 305–310.
- Connelly D.P., Copley J.T., Murton B.J., Stansfield K., Tyler P.A., German C.R., Van Dover C.L., Amon D., Furlong M., Grindlay N., Hayman N., Hühnerbach V., Judge M., Le Bas T., McPhail S., Meier A., Nakamura K.-I., Nye V., Pebody M., Pedersen R.B., Plouviez S., Sands C., Searle R.C., Taws S. and Wilcox S. (in press) Hydrothermal vent fields and chemosynthetic biota on the world's deepest seafloor spreading centre. *Nature Communications*.
- Dana J.D. (1852) *Crustacea. Part I. United States Exploring Expedition. During the years 1838, 1839, 1840, 1841, 1842. Under the command of Charles Wilkes, U.S.N., Volume 13.* Philadelphia: C. Sherman.
- De Grave S., Pentcheff N.D., Ahyong S.T., Chan T.Y., Crandall K.A., Dworschak P.C., Felder D.L., Feldmann R.M., Franssen C.H.J.M., Goulding L.Y.D., Lemaitre R., Low M.E.Y., Martin J.W., Ng P.K.L., Schweitzer C.E., Tan S.H., Tshudy D. and Wetzer R. (2009) A classification of living and fossil genera of decapod crustaceans. *Raffles Bulletin of Zoology Supplement* 21, 1–109.
- Desbruyères D., Segonzac M. and Bright M. (2006) *Handbook of deep-sea hydrothermal vent fauna.* Vienna: Biologiezentrum der Oberösterreichische Landesmuseen.
- Doyle J.J. and Dickson E. (1987) Preservation of plant samples from DNA restriction endonuclease analysis. *Taxon* 36, 715–722.
- Escobar-Briones E. and Villalobos Hiriart J.L. (2003) Deep-water caridean shrimps (Crustacea: Decapoda) from Banco Chinhorro and adjacent areas in the Northern Caribbean. *Bulletin of Marine Science* 73, 99–122.
- Fabri M.-C., Bargain A., Briand P., Gebruk A., Fouquet Y., Morineaux M. and Desbruyères D. (2011) The hydrothermal vent community of a new deep-sea site, Ashadze-1, 12°58'N on the Mid-Atlantic Ridge. *Journal of the Marine Biological Association of the United Kingdom* 91, 1–13.
- Folmer O., Black M., Hoeh W., Lutz R. and Vrijenhoek R. (1994) DNA primers for amplification of mitochondrial cytochrome c oxidase subunit I from diverse metazoan invertebrates. *Molecular Marine Biology and Biotechnology* 3, 294–299.
- Fujikura K., Hashimoto J., Fujiwara Y. and Okutani T. (1995) Community ecology of the chemosynthetic community at Off Hatsushima site, Sagami Bay, Japan. *JAMSTEC Journal of Deep-Sea Research* 11, 227–241.
- Gebruk A.V., Pimenov N.V. and Savvichev A.S. (1993) Feeding specialization of bresiliid shrimps in the TAG site hydrothermal community. *Marine Ecology Progress Series* 98, 247–253.
- Gebruk A.V., Galkin S.V., Vereshchaka A.L., Moskalev L.I. and Southward A.J. (1997) Ecology and biogeography of the hydrothermal vent fauna of the Mid-Atlantic Ridge. *Advances in Marine Biology* 32, 93–144.
- German C.R., Bowen A., Coleman M.L., Honig D.L., Huber K.A., Jakuba M. V., Kinsey J.C., Kurz M.D., Leroy S., McDermott J.M., Mercier de Lepinay B., Nakamura K., Seewald J.S., Smith J. L., Sylva S.P., Van Dover C.L., Whitcomb L.L. and Yoerger D.R. (2010) Diverse styles of submarine venting on the ultraslow spreading Mid-Cayman Rise. *Proceedings of the National Academy of Sciences of the United States of America* 107, 14020–14025.
- Hall T.A. (1999) BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. *Nucleic Acids Symposium Series* 41, 95–98.
- Hashimoto J., Ohta S., Gamo T., Chiba H., Yamaguchi T., Tsuchida S., Okudaira T., Watabe H., Yamanaka T. and Kitazawa M. (2001) First hydrothermal vent communities from the Indian Ocean discovered. *Zoological Science* 18, 717–721.
- Holthuis L.B. (1993) *The recent genera of the caridean and stenopodidean shrimps (Crustacea, Decapoda) with an appendix on the order Amphionidacea.* Leiden: Nationaal Natuurhistorisch Museum.
- Jukes T.H. and Cantor C.R. (1969) Evolution of protein molecules. In Munro H.N. (ed.) *Mammalian protein metabolism.* New York: Academic Press, pp. 21–132.
- Kikuchi T. and Ohta S. (1995) Two caridean shrimps of the families Bresiliidae and Hippolytidae from a hydrothermal field on the Iheya-Ridge, off the Ryukyu-Islands, Japan. *Journal of Crustacean Biology* 15, 771–785.
- Kikuchi T. and Hashimoto J. (2000) Two new caridean shrimps of the family Alvinocarididae (Crustacea, Decapoda) from a hydrothermal knoll in the Mid-Okinawa Trough, Japan. *Species Diversity* 5, 135–148.
- Kimura M. (1980) A simple method for estimating evolutionary rate of base substitutions through comparative studies of nucleotide sequences. *Journal of Molecular Evolution* 16, 111–120.
- Komai T. and Segonzac M. (2003) Review of the hydrothermal vent shrimp genus *Mirocaris*, redescription of *M. fortunata* and reassessment of the taxonomic status of the family Alvinocarididae (Crustacea: Decapoda: Caridea). *Cahiers de Biologie Marine* 44, 199–215.
- Komai T. and Segonzac M. (2004) A new genus and species of alvinocaridid shrimp (Crustacea: Decapoda: Caridea) from hydrothermal vents on the North Fiji and Lau Basins, south-western Pacific. *Journal of the Marine Biological Association of the United Kingdom* 84, 1179–1188.

- Komai T. and Segonzac M.** (2005) A revision of the genus *Alvinocaris* Williams and Chace (Crustacea: Decapoda: Caridea: Alvinocarididae), with descriptions of a new genus and a new species of *Alvinocaris*. *Journal of Natural History* 39, 1111–1175.
- Komai T. and Segonzac M.** (2006) *Chorocaris chacei* Williams & Rona, 1986, *Chorocaris paulexa* Martin & Shank, 2005, *Chorocaris vandoverae* Martin & Hessler, 1990, *Rimicaris exoculata* Williams & Rona, 1986. In Desbruyères D., Segonzac M. and Bright M. (eds) *Handbook of deep-sea hydrothermal vent fauna*. Vienna: Biologiezentrum der Oberösterreichische Landesmuseen, pp. 421–423.
- Komai T. and Segonzac M.** (2008) Taxonomic reviews of the hydrothermal vent shrimp genera *Rimicaris* Williams & Rona and *Chorocaris* Martin & Hessler (Crustacea: Decapoda: Caridea: Alvinocarididae). *Journal of Shellfish Research* 27, 21–41.
- Komai T. and Chan T.Y.** (2010) A new genus and two new species of alvinocaridid shrimps (Crustacea: Decapoda: Caridea) from a hydrothermal vent field off northwestern Taiwan. *Zootaxa* 2372, 15–32.
- Komai T. and Yamada Y.** (2010) A new species of the rare caridean genus *Bresilia* Calman (Crustacea: Decapoda: Bresiliidae) from the Ryukyu Islands, Japan, representing a family new to the North Pacific marine fauna. *Zootaxa* 2450, 41–52.
- Komai T. and Yamada Y.** (2011) A new species of the caridean genus *Bresilia* Calman (Decapoda: Bresiliidae) discovered from a shallow-water submarine cave in Okinawa Islands, Japan. *Bulletin of the National Museum of Nature and Science Series A Supplement* 5, 71–82.
- Komai T., Shank T.M. and Van Dover C.L.** (2005) A new species of *Alvinocaris* (Crustacea: Decapoda: Caridea: Alvinocarididae) and a new record of *A. muricola* from methane seeps on the Blake Ridge Diapir, north-western Atlantic. *Zootaxa* 1019, 27–42.
- Komai T., Martin J.W., Zala K., Tsuchida S. and Hashimoto J.** (2006) A new species of *Mirocaris* (Crustacea: Decapoda: Caridea: Alvinocarididae) associated with hydrothermal vents on the Central Indian Ridge, Indian Ocean. *Scientia Marina* 70, 109–119.
- Komai T., Giere O. and Segonzac M.** (2007) New record of alvinocaridid shrimps (Crustacea: Decapoda: Caridea) from hydrothermal vent fields on the southern Mid-Atlantic Ridge, including a new species of the genus *Opaepele*. *Species Diversity* 12, 237–253.
- Kuenzler R.O., Kwasniewski J.T., Jinks R.N., Lakin R.C., Battelle B. -A., Herzog E.D., Renninger G.H. and Chamberlain S.C.** (1997) Retinal anatomy of new bresiliid shrimp from the Lucky Strike and Broken Spur hydrothermal vent fields on the Mid-Atlantic Ridge. *Journal of the Marine Biological Association of the United Kingdom* 77, 707–725.
- Lakin R.C., Jinks R.N., Battelle B.-A., Herzog E.D., Kass L., Renninger G.H. and Chamberlain S.C.** (1997) Retinal anatomy of *Chorocaris chacei*, a deep-sea hydrothermal vent shrimp from the Mid-Atlantic Ridge. *Journal of Comparative Neurology* 383, 503–514.
- Latreille P.A.** (1802) *Histoire naturelle, générale et particulière des Crustacés et des Insectes. Ouvrage faisant suite à l'histoire naturelle générale et particulière, composée par Leclerc de Buffon, et rédigée par C.S. Sonnini, membre de plusieurs sociétés savantes. Familles naturelles des genres, Volume 3*. Paris: F. DuFart.
- Lunina A.A. and Vereshchaka A.L.** (2010) A new vent shrimp (Crustacea: Decapoda: Alvinocarididae) from the Mid-Atlantic Ridge. *Zootaxa* 2372, 69–74.
- Martin J.W. and Hessler R.R.** (1990) *Chorocaris vandoverae*, a new genus and species of hydrothermal vent shrimp (Crustacea: Decapoda: Bresiliidae) from hydrothermal vent fields along the Mid-Atlantic Ridge. *Contributions in Science, Natural History Museum of Los Angeles County* 417, 1–11.
- Martin J.W. and Christiansen J.C.** (1995) A new species of the shrimp genus *Chorocaris* Martin and Hessler, 1990 (Crustacea, Decapoda, Bresiliidae) from hydrothermal vent fields along the Mid-Atlantic Ridge. *Proceedings of the Biological Society of Washington* 108, 220–227.
- Martin J.W. and Davis G.E.** (2001) An updated classification of the Recent Crustacea. *Natural History Museum of Los Angeles County, Science Series* 39, 1–124.
- Martin J.W. and Haney T.A.** (2005) Decapod crustaceans from hydrothermal vents and cold seeps: a review through 2005. *Zoological Journal of the Linnean Society* 145, 445–522.
- Martin J.W. and Shank T.M.** (2005) A new species of the shrimp genus *Chorocaris* (Decapoda: Caridea: Alvinocarididae) from hydrothermal vents in the eastern Pacific Ocean. *Proceedings of the Biological Society of Washington* 111, 183–198.
- Martin J.W., Signorovich J. and Patel H.** (1998) Comparison of the carpal cleaning brush in two genera of hydrothermal vent shrimp (Crustacea, Decapoda, Bresiliidae). *Journal of Morphology* 235, 31–39.
- Ohta S. and Kim D.** (2001) Submersible observations of the hydrothermal vent communities on the Iheya Ridge, Mid Okinawa Trough, Japan. *Journal of Oceanology* 57, 663–667.
- O'Neill P.J., Jinks R.N., Herzog E.D., Battelle B.-A., Kass L., Renninger G.H. and Chamberlain S.C.** (1995) The morphology of the dorsal eye of the hydrothermal vent shrimp, *Rimicaris exoculata*. *Visual Neuroscience* 12, 861–875.
- Palumbi S.R.** (1996) Nucleic acids II: the polymerase chain reaction. In Hillis D.M., Moritz C. and Mable B.K. (eds) *Molecular systematics*. Sunderland, MA: Sinauer Associates, pp. 205–247.
- Peli D.G. and Chamberlain S.C.** (1989) The visibility of 350°C blackbody radiation by the shrimp *Rimicaris exoculata* and man. *Nature* 337, 460–461.
- Rosencrantz E., Ross I.R. and Sclater J.G.** (1988) Age and spreading history of the Cayman Trough as determined from depth, heat flow, and magnetic anomalies. *Journal of Geophysical Research* 93, 2141–2157.
- Saitou N. and Nei M.** (1987) The neighbour-joining method: a new method for reconstructing phylogenetic trees. *Molecular Biology and Evolution* 4, 406–425.
- Segonzac M.** (1992) The hydrothermal vent communities of Snake Pit area (Mid-Atlantic Ridge, 23°N, 3,480 m)—megafaunal composition and microdistribution. *Comptes Rendus de l'Académie des Sciences* 314, 593–600.
- Segonzac M., de Saint Laurent M. and Casanova B.** (1993) L'enigme du comportement trophiques des crevettes Alvinocarididae des sites hydrothermaux de la dorsale médio-atlantique. *Cahiers de Biologie Marine* 34, 535–571.
- Shank T.M. and Martin J.W.** (2003) A new caridean shrimp of the family Alvinocarididae from thermal vents at Menez Gwen on the Mid-Atlantic Ridge. *Proceedings of the Biological Society of Washington* 116, 158–167.
- Shank T.M., Black M.B., Halanych K.M., Lutz R.A. and Vrijenhoek R.C.** (1999) Miocene radiation of deep-sea hydrothermal vent shrimp (Caridea: Bresiliidae): evidence from mitochondrial cytochrome oxidase subunit I. *Molecular Phylogenetics and Evolution* 13, 244–254.
- Sogin M.L.** (1990) Amplification of ribosomal RNA genes for molecular evolution studies. In Innis M.A., Gelfand D.H., Sninsky J.J. and White T.J. (eds) *PCR protocols. A guide to methods and applications*. San Diego, CA: Academic Press, pp. 307–314.
- Tamura K., Peterson D., Peterson N., Stecher G., Nei M. and Kumar S.** (2011) MEGA5: Molecular Evolutionary Genetics Analysis using Maximum Likelihood, Evolutionary Distance, and Maximum Parsimony Methods. *Molecular Biology and Evolution* 28, 2731–2739. doi:10.1093/molbev/msr121.

- Tsuchida S., Yamaguchi T., Komai T and Watanabe H.** (2008) Arthropoda. In Fujikura K., Okutani T. and Maruyama T. (eds) *Deep-sea life—biological observations using research submersibles*. Hatano: Tokai University Press, pp.100–178.
- Van Dover C.L.** (2000) *The ecology of deep-sea hydrothermal vents*. Princeton: Princeton University Press.
- Van Dover C.L., Fry B., Grassle J.F., Humphris S. and Rona P.A.** (1988) Feeding biology of the shrimp *Rimicaris exoculata* at hydrothermal vents on the Mid-Atlantic Ridge. *Marine Biology* 98, 209–216.
- Van Dover C.L., Szuts E.Z., Chamberlain S.C. and Cann J.R.** (1989) A novel eye in eyeless shrimp from hydrothermal vents on the Mid-Atlantic Ridge. *Nature* 337, 458–460.
- Van Dover C.L., Cann J.R., Cavanaugh C., Chamberlain S., Delaney J.R., Janecky D., Imhoff J. and Tyson J.A.** (1994) Light at deep sea hydrothermal vents. *EOS, Transactions of the American Geophysical Union* 75, 44–45.
- Van Dover C.L., Reynolds G.T., Chave A.D. and Tyson J.A.** (1996) Light at deep-sea hydrothermal vents. *Geophysical Research Letters* 23, 2049–2052.
- Van Dover C.L., Humphris S.E., Fornari D., Cavanaugh C.M., Collier R., Goffredi S.K., Hashimoto J., Lilley M.D., Reysenbach A.L., Shank T.M., Von Damm K.L., Banta A., Gallant R.M., Gotz D., Green D., Hall J., Harmer T.L., Hurtado L.A., Johnson P., McKiness Z.P., Meredith C., Olson E., Pan I.L., Turnipseed M., Won Y., Young C.R. and Vrijenhoek R.C.** (2001) Biogeography and ecological setting of Indian Ocean hydrothermal vents. *Science* 294, 818–823.
- Van Dover C.L., Aharon P., Bernhard J.M., Caylor E., Doerries M., Flickinger W., Gilhooly W., Goffredi S.K., Knick K.E., Macko S.A., Rapoport S., Raulfs E. C., Ruppel C., Salerno J.L., Seitz R.D., Sen Gupta B.K., Shank T., Turnipseed M. and Vrijenhoek R.** (2003) Blake Ridge methane seeps: characterization of a soft-sediment, chemosynthetically based ecosystem. *Deep-Sea Research I* 50, 281–300.
- Vereshchaka A.L.** (1996) A new genus and species of caridean shrimp (Crustacea: Decapoda: Alvinocarididae) from North Atlantic hydrothermal vents. *Journal of the Marine Biological Association of the United Kingdom* 76, 951–961.
- Vereshchaka A.L.** (1997) A new family for a deep-sea caridean shrimp from North Atlantic hydrothermal vents. *Journal of the Marine Biological Association of the United Kingdom* 77, 245–438.
- Watabe H. and Miyake H.** (2000) Decapod fauna of the hydrothermally active and adjacent fields on the Hatoma Knoll, southern Japan. *JAMSTEC Journal of Deep Sea Research* 17, 29–34.
- Watabe H. and Hashimoto J.** (2002) A new species of the genus *Rimicaris* (Alvinocarididae: Caridea: Decapoda) from the active hydrothermal vent field, 'Kairei Field,' on the Central Indian Ridge, the Indian Ocean. *Zoological Science* 19, 1167–1174.
- Webber W.R.** (2004) A new species of *Alvinocaris* (Crustacea: Decapoda: Alvinocarididae) and new records of alvinocaridids from hydrothermal vents north of New Zealand. *Zootaxa* 444, 1–26.
- Wicksten M.K.** (1989) *Encantada spinocolata*, a new genus and species of shrimp from the Galapagos Islands (Caridea, Bresiliidae). *Journal of Crustacean Biology* 9, 667–671.
- Williams A.B.** (1988) New marine decapod crustaceans from waters influenced by hydrothermal discharge, brine and hydrocarbon seepage. *Fishery Bulletin* 86, 263–287.
- Williams A.B. and Chace F.A. Jr** (1982) A new caridean shrimp of the family Bresiliidae from thermal vents of the Galapagos Rift. *Journal of Crustacean Biology* 2, 136–147.
- Williams A.B. and Rona P.A.** (1986) Two new caridean shrimps (Bresiliidae) from a hydrothermal field on the Mid-Atlantic Ridge. *Journal of Crustacean Biology* 6, 446–462.
- Williams A.B. and Dobbs F.C.** (1995) A new genus and species of caridean shrimp (Crustacea: Decapoda: Bresiliidae) from hydrothermal vents on Loihi Seamount, Hawaii. *Proceedings of the Biological Society of Washington* 108, 227–237.
- and
- Zelnio K.A. and Hourdez S.** (2009) A new species of *Alvinocaris* (Crustacea: Decapoda: Caridea: Alvinocarididae) from hydrothermal vents at the Lau Basin, southwest Pacific, and a key to the species of Alvinocarididae. *Proceedings of the Biological Society of Washington* 122, 52–71.

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