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**PREY SPECTRA OF PLEURONECTIDS
(HIPPOGLOSSOIDES PLATESSOIDES,
PLEURONECTES FERRUGINEUS,
PLEURONECTES AMERICANUS) FROM
SABLE ISLAND BANK**

by:

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PREY SPECTRA OF PLEURONECTIDS (Hippoglossoides platessoides, Pleuronectes ferrugineus, Pleuronectes americanus) FROM SABLE ISLAND BANK

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ABSTRACT

Martell, D.J. and G. McClelland. 1992. Prey spectra of pleuronectids (Hippoglossoides platessoides, Pleuronectes ferrugineus, Pleuronectes americanus) from Sable Island Bank. Can. Tech. Rep. Fish. Aquat. Sci. 1895: vi + 20p.

Diets were documented for Hippoglossoides platessoides, Pleuronectes ferrugineus, and Pleuronectes americanus collected from southwest Sable Island Bank in February and June 1989. Diets varied seasonally and with species and length of fish. Of 239 species of prey consumed by flatfishes, 66 were found in the stomachs of at least 5% of each fish species sampled, frequently enough to be considered principal prey. Crustaceans, especially amphipods, were the prey most frequently exploited by all three flatfish species. Small Hippoglossoides platessoides fed on suprabenthic fauna, while larger fish exploited epifauna and hyperiid suprafaua. Pleuronectes ferrugineus exploited epibenthic fauna, with crustaceans and tunicates being consumed by smaller and polychaetes by larger fish. Pleuronectes americanus also preyed upon epifauna, although the smaller individuals exploited polychaetes and larger fish, crustaceans associated with ectoproct colonies. Feeding intensity declined dramatically in winter, food being found in the stomachs of only 36% of H. platessoides and 49% of P. ferrugineus sampled in February; stomachs of all P. americanus collected in February were empty. In contrast, 86% of H. platessoides, 94% of P. ferrugineus, and 96% of P. americanus sampled in June had food in their stomachs.

RÉSUMÉ

Martell, D.J. et G. McClelland. 1992. Spectre de proies pleuronectidés (Hippoglossoides platessoides, Pleuronectes ferrugineus, Pleuronectes americanus) du banc de l'Île de Sable. Can. Tech. Rep. Fish. Aquat. Sci. 1895: vi + 20p.

On a documenté le régime alimentaire d'Hippoglossoides platessoides, de Pleuronectes ferrugineus et de Pleuronectes americanus prélevés dans le sud-ouest du banc de l'Île de Sable en février et en juin 1989. Le régime a varié selon la saison et selon l'espèce et la longueur des poissons. Des 239 espèces de proies consommées par les poissons plats, 66 ont été retrouvées dans l'estomac d'au moins 5% de chacune des espèces échantillonnées, soit une fréquence suffisante pour qu'elles soient considérées comme des proies principales. Les crustacés, et surtout les amphipodes, étaient les proies les plus fréquemment exploitées par les trois espèces de poissons plats. Les Hippoglossoides platessoides de petite taille se nourrissaient de la faune suprabenthique, tandis que les poissons de plus grande taille exploitaient l'épifaune et la suprafaua des hyperiidés. Les Pleuronectes ferrugineus exploitaient la faune épibenthique, les représentants de plus petite taille consommant des crustacés et des tuniqueés et les représentants de plus grande taille, des polychètes. Les Pleuronectes americanus s'alimentaient également au dépens de l'épifaune, cette fois les individus de petite taille exploitant les polychètes et les poissons de plus grande taille, des crustacés associés aux colonies d'ectoproctes. L'intensité de l'alimentation a diminué de façon très marquée en hiver, des aliments ayant été observé chez seulement 36% des H. platessoides et 49% des P. ferrugineus échantillonnés en février; tous les P. americanus échantillonnés en février avaient l'estomac vide. Par comparaison, 86% des H. platessoides, 94% des P. ferrugineus et 96% des P. americanus échantillonnés en juin avaient de la nourriture dans l'estomac.

INTRODUCTION

As part of a larger study of larval sealworm (*Pseudoterranova decipiens*) transmission to demersal fish on Sable Island Bank, a comparative survey of the diets of three common flatfish species was undertaken to provide indirect, if not direct, evidence leading to the identification of important invertebrate precursor hosts of the parasite (McClelland *et al.* 1990). Flatfish (Pleuronectiformes) found off Canada's east coast are generally considered to be benthic invertebrate predators (Grosslein *et al.* 1980, Hacunda 1981, Langton and Bowman 1981, Scott and Scott 1988). Diet studies have indicated that, at a broad taxonomic level, several of the common species of flatfish found on Sable Island Bank exploit similar prey (Scott 1975a, 1975b, 1976). This report presents the complete spectra of prey consumed by American plaice, yellowtail flounder, and winter flounder from southwestern Sable Island Bank. Statistical analyses of variations in abundances of principal prey in flatfish diets and identifications of invertebrate hosts of *P. decipiens* among flatfish prey are found in a thesis (Martell 1992) and will be dealt with in the future in primary literature.

MATERIALS AND METHODS

Samples of three species of flatfish, American plaice (*Hippoglossoides platessoides*), yellowtail flounder (*Pleuronectes ferrugineus*), and winter flounder (*Pleuronectes americanus*), were collected from a 1,870 square kilometre (km²) area on Sable Island Bank (Fig. 1). Fish were taken from a series of replicated sets using a Western IIA otter trawl with a fine-mesh (1 cm) codend liner during Department of Fisheries and Oceans research cruises, on the "Alfred Needler", February 10-18, 1989 and on the "Lady Hammond", June 5-15, 1989. All tows were 30 minutes in duration and made at a speed of 6.5 kilometres per hour.

Fish were chosen randomly from each tow and were stratified into seven 5 cm length groups (i.e., ≤15, 16-20, 21-25, 26-30, 31-35, 36-40, ≥41 cm) each containing a maximum of 30 fish (McClelland *et al.* 1983). Total length (TL) and sex of each fish were recorded and,

after the stomach was removed, the body was tagged and frozen. The stomach was then placed in a 70% ethanol solution, and labeled with the tag number. Fish ≤15 cm TL were measured, sexed, tagged, and preserved whole in 70% ethanol with an incision in the abdominal wall to expose the viscera. On return to the laboratory, the contents of each stomach were removed, sorted, identified to species, when possible, and counted. Prey were identified using the following keys: Smith 1964, Barnard 1969, Shultz 1969, Gosner 1971, Bousfield 1973, Bowman 1973, Barnes 1974, Jones 1976, Lincoln 1979, Appy *et al.* 1980, Holdich and Jones 1983, Schneppenheim and Weigmann-Haass 1986, Scott and Scott 1988, Squires 1990.

For each flatfish species, prey occurrences were determined for only those fish with food in their stomachs. Prey species were stratified into <5%, 5-49%, and ≥50% levels of occurrence, with principal prey determined to have an occurrence of ≥5%.

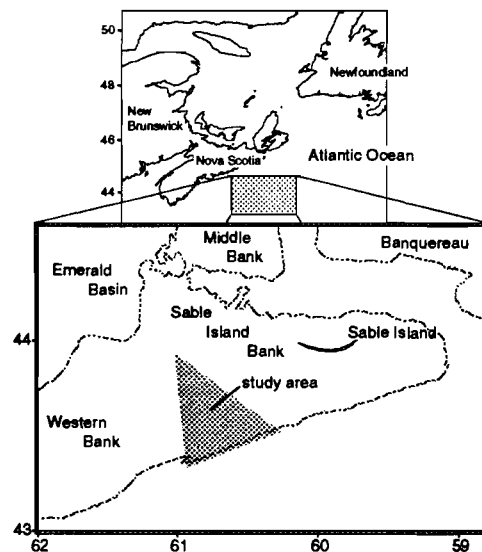


Fig. 1. Sampling area for flatfish diet study in February and June 1989.

RESULTS

A total of 801 flatfish were sampled, 387 in February and 414 in June 1989. Food was found in the stomachs of 147 (38%) of the fish collected in winter and in the stomachs of 379 (92%) of those sampled in summer (Table 1). Of

239 prey species, 66 consumed by $\geq 5\%$ of each of those flatfish species that had food in their stomachs, were considered principal prey (Table 2).

Of 337 American plaice (*Hippoglossoides platessoides*) sampled, 183 specimens were taken in February and 154 in June. Although 100 species of prey were consumed by American plaice (52% crustaceans and 29% polychaetes), only 18 qualified as principal prey (Table 2). The most frequently exploited crustaceans were cumaceans, hyperiid and gammaridean amphipods, mysids, and pagurids (Tables 3 & 4). Anthozoans, molluscs, and echinoderms were also common prey. Food was absent in stomachs of 118 (64% of 183) of the plaice sampled in February while only 22 (14%) of those taken in June were empty (Table 1). During winter there was also a decrease in both diversity and quantity of prey consumed.

Yellowtail flounder (*Pleuronectes ferrugineus*) collected included 169 and 141 specimens from February and June respectively, for a total of 310. Of 239 species of prey consumed by the three flatfish species, 184 were exploited by yellowtail (Table 2). Yellowtail fed primarily upon polychaetes (83 species including phyllodocids, polynoids, spionids, orbinids, cirratulids, and ampharetids), crustaceans (68 species with cumaceans and gammaridean amphipods predominant), and one species of tunicate (Tables 3 & 4). Overall, 43 (23%) species were principal prey and 3 (2%) species had been consumed by $\geq 50\%$ of the yellowtail that had food in their stomachs. Stomachs of 85 (51%) of the yellowtail flounder collected in February were empty while prey were absent in stomachs of only 8 (6%) of those taken in June (Table 1). Similar to plaice, yellowtail consumed fewer species and lesser quantities of prey in February than in June.

Winter flounder (*Pleuronectes americanus*) specimens numbered 177, 47 collected in February and 130 in June. Of 140 species of prey exploited by winter flounder (Table 2), 67 (48%) were polychaetes, such as phyllodocids, spionids, and terebellids and 52 (37%) were crustaceans, including cumaceans and gammaridean and caprellid amphipods, and one, a species of tunicate (Tables 3 & 4).

Forty species (29%) were principal prey but only one species of prey occurred in $\geq 50\%$ of stomachs that contained food. Stomachs of all fish sampled in February were empty, although food was present in the intestines of some specimens. In contrast, 116 (96%) of the fish collected in June had food in their stomachs (Table 1).

DISCUSSION

In the present study a wide variety of prey was consumed by American plaice (*Hippoglossoides platessoides*), yellowtail flounder (*Pleuronectes ferrugineus*), and winter flounder (*Pleuronectes americanus*). Although the diets of these fish appeared to be superficially similar, closer inspection revealed differences in prey selection and, by inference, feeding behavior. In this study flatfish diets often consisted of polychaetes and crustaceans, with crustaceans, especially amphipods, being the most common principal prey (Tables 3 & 4). Crustaceans consumed included 19 species of amphipods, 17 of which were gammarideans. Coull and Bell (1983) report that sandy bottom invertebrate communities are dominated by crustaceans and that amphipods make up the majority of the macrofauna. Wildish *et al.* (1989), however, reported that polychaete species outnumbered crustacean species by 2.5 fold in grab samples of similar substrate (Sable Island sand and gravel) from Brown's Bank. Other diet studies have also revealed that crustaceans, particularly amphipods, were major components of flatfish diets (Powles 1965, Minet 1973, Hacunda 1981, Langton and Bowman 1981, Wakabara *et al.* 1982, Langton 1983, Hahn and Langton 1984, Worobec 1984, Macdonald and Green 1986) and were also prominent in the diets of other demersal fish species (Tyler 1972, Kislalioglu and Gibson 1977, Grosslein *et al.* 1980, Hacunda 1981, Gibson and Ezzi 1987).

The most obvious variation in the diets of the three flatfish species from Sable Island Bank in the present study was the seasonal disparity in feeding intensity (Table 1). Similar declines in feeding intensity have been reported in studies of American plaice in the Gulf of St. Lawrence (Powles 1965), St. Margarets Bay, N.S. (MacKinnon 1972), St.

Pierre Bank (Minet 1973), the Grand Banks (Pitt 1973), and the Bay of Fundy (Macdonald and Green 1986). They have also been documented for yellowtail flounder off Cape Ann (Libey and Cole 1979), northeastern North American offshore (Langton 1983), Georges Bank (Collie 1987). Winter flounder in Long Pond, Conception Bay, Newfoundland (Kennedy and Steele 1971), St. Margarets Bay (Levings 1974), Gulf of Maine (Hacunda 1981), Passamaquoddy Bay (McLeese and Moon 1989), and various demersal marine fish species from the North Atlantic (Dawson and Grimm 1980, Hacunda 1981, Macdonald and Green 1986) also consume less food during winter. The decline in feeding intensity in February may have coincided with a reduction in the abundance of marine invertebrate fauna which typically occurs in northern benthic communities in winter as reported in studies of St. Margarets Bay, Nova Scotia (MacKinnon 1972), New England salt pond (Worobec 1984), and Long Island Sound (Birnbaum 1979). Wildish *et al.* (1989), however, reported that a seasonal decline in total benthic production did not occur on Brown's Bank, an area with sediments identical to those found on Sable Island Bank (i.e., Sable Island sand and gravel) (King 1970, Fader *et al.* 1977). The apparent lack of such a decline in Wildish *et al.* (1989), however, may have been attributable to high sample variances.

The reduction in numbers and variety of invertebrates in winter diets of both plaice and yellowtail may have been indicative of a decline in availability of benthic fauna, as fish diets are reported to reflect the habitats that they exploit for food (McEachran *et al.* 1976, Ringler 1979, Stoner 1979, Macdonald and Green 1986, Delbeek and Williams 1988). However, prey availability for a predator is not only a function of the presence or absence of prey but also of prey size, behavior, and density within an exploited habitat (Moore and Moore 1976, Ringler 1979). All of these criteria act together to determine how a predator will select its food resources (Moore and Moore 1976). If, within a fishes' habitat, some available prey are too small or too large, inactive, or sparsely distributed, changes in diet may occur (Moore and Moore 1976, Ringler 1979). Such variations may also result from a change in prey habitats exploited by the fish (Jones 1952, Langton 1983,

Macdonald and Green 1986, Collie 1987) although, in the present study, fish were collected, over identical habitats (King 1970).

Principal prey, which Hacunda (1981) defined as a primary food resource exploited by a fish, included 64 invertebrate and 2 chordate taxa in the present study. As other authors (Tyler 1972, Macdonald and Green 1983) have found, only a few of the prey species in each fish species' diet were principal prey. In the present study, American plaice consumed 18 species of principal prey, while yellowtail flounder and winter flounder exploited 43 and 41 principal prey species, respectively. Plaice appeared to specialize upon crustaceans while the other two flatfish species had diets more typical of generalists (Schoener 1971, Crow 1978).

Prey were assigned to three functional classifications or guilds, infauna, epifauna, and suprabenthic fauna (Martell 1992). These categories were established according to the habitat requirements, feeding patterns, and activity levels of prey species (Barnes 1969, Bousfield 1973, Birnbaum 1979, Fauchald and Jumars 1979, Sainte-Marie and Brunel 1985, Wildish *et al.* 1992). The infaunal guild, composed of prey found free or living in tubes in the sediment or, at times venturing above the sediment to feed, may be deposit- (eg. the polychaete *Aricidea suecica*) or filter-feeders (eg. the amphipod *Ampelisca macrocephala*). Epifauna, live and feed at the sediment surface and may be found in tubes attached to a substrate (eg. the amphipod *Ischyroceros anguipes*), clinging to a substrate (eg. the caprellid *Aeginina longicornis*), or moving freely about the sediment surface (eg. the brittle star *Amphipholis squamata*). Some epifauna (eg. the sea anemone *Actinauge longicornis*, ectoprocts) are sedentary, attaching themselves to a substrate. Suprabenthic fauna (eg. cumaceans, hyperiid amphipods, and mysids), maintain a requisite association with the benthos, but also undertake vertical migrations of varying magnitudes (Sainte-Marie and Brunel 1985, Wildish *et al.* 1992).

Trophic classifications of polychaetes and amphipods according to their habitat requirements and feeding have been well documented. Fauchald and Jumars (1979)

grouped polychaete taxa into several guilds, which may be further categorized into four basic types, the mobile carnivores (eg. Nephtys sp.), the burrowers (eg. Ophelina acuminata), the surface deposit-feeders (eg. Ampharete sp.), and the filter-feeders (eg. Exogone hebes) (Martell 1992). Information describing basic lifestyles and habitat requirements of gammaridean amphipods (Bousfield 1973, Biernbaum 1979) and criteria established by other investigators (Sainte-Marie and Brunel 1985, Wildish et al. 1992, Mills pers. comm.) were used here to create the free-swimming (eg. Stenopleustes inermis), burrowing (eg. Monoculodes edwardsi), nestling (Biernbaum 1979) (eg. Photis sp.), and tubaceous (infaunal and epifaunal) (eg. Erichthonius rubricornis) amphipod guilds (Martell 1992).

Diets of plaice, herein, showed variation with fish length only during summer, but trends in prey guild exploitation were apparent from the diets of February and June specimens. The diets of the smallest fish (<21 cm TL) were composed of suprabenthic (eg. mysids) and epibenthic crustaceans (eg. tubaceous amphipods) and, but for the inclusion of hyperiid amphipods during summer (Tables 3 & 4), were similar to those of small plaice from the southern Gulf of St. Lawrence (Powles 1965). Suprafauna, with the addition of hyperiid amphipods during summer, were again prominent in the diets of fish 21-30 cm TL, although mysid exploitation declined for larger fish of this length stratum. In winter, large fish (≥ 31 cm TL) usually consumed epibenthic prey of a sedentary (eg. Actinauge longicornis) or slow-moving (eg. Pagurus sp.) nature but, like the smaller fish, frequently preyed upon suprafauna, such as hyperiids, during summer. Plaice appear to progress through two feeding stanzas, exploiting suprafauna when small and larger epifauna as they matured.

In the present study, small yellowtail flounder (≤ 30 cm TL) consumed epifauna, consisting primarily of cumaceans and some amphipods during both winter and summer (Tables 3 & 4). In winter, tubaceous amphipods, such as corophids and aorids, were common prey, although consumption of most of these crustaceans declined in larger fish. Increased fish length was accompanied by greater

exploitation of epifauna (echinoderms), infauna (polychaetes and amphipods). Libey and Cole (1979) also found that cumaceans were a major component of yellowtail flounder diets, but they failed to document variations in prey consumption with fish length. Despite being included among suprabenthic fauna, cumaceans are filter-feeders that often inhabit the top layer of sediment (Barnes 1974, Jones 1976). Amphipods (Langton 1983) and, specifically, tubaceous forms (eg. Unciola irrorata) (Collie 1987) are often identified as common prey of small yellowtail flounder. Hacunda (1981) reported that yellowtail foraged at the sediment surface, consuming epifauna and available infauna (usually polychaetes) and, perhaps as Libey and Cole (1979) suggest, ingesting the upper sediment layers to find food. Collie (1987) also related that yellowtail often consumed portions of epifaunal prey such as the tentacles of polychaetes which were exposed at the sediment surface.

Large yellowtail consume suprabenthic fauna, such as Mysis mixta and Crangon septemspinosa (Hacunda 1981, Langton 1983, Collie 1987). In the present study, large yellowtail exploited cumaceans and mysids in winter, but, in summer, they fed mainly on infaunal polychaetes, particularly surface deposit-feeders (eg. Spiophanes bombyx) (also found by Hacunda 1981), and burrowers (eg. Ophelina acuminata) and also exploited epifaunal polychaetes (eg. Phyllodoce sp.), tunicates (Molgula sp.), and tubaceous amphipods. Yellowtail of Sable Island Bank seemed to progress through two feeding stanzas, exploiting small epifauna or infauna when small and larger infauna (polychaetes and tubaceous amphipods) as they matured.

Winter flounder, in the present study, primarily exploited epifauna, including cumaceans which, as Kennedy and Steele (1971) pointed out, were probably epi- or infaunal when consumed. Small surface deposit-feeding polychaetes (eg. Ampharete lindstroemi) and nestling (eg. Metopa bruzelli) and free-swimming amphipods (eg. Pontogeneia inermis) were also ingested but were replaced by tubaceous amphipods in the diets of larger fish (Tables 3 & 4). As Worobec (1984) also observed, epifauna ingested by larger fish were probably consumed incidentally with tunicates.

Polychaetes consumed by large winter flounder in this study and in a Gulf of Maine study (Hacunda 1981), were mobile epifaunal carnivores, such as Phyllodoce sp. This group of winter flounder consumed greater quantities of ectoprocts and associated tubaceous amphipods (Erichthonius rubricornis and Ischyroceros anguipes) and clinging caprellids (Aeginina longicornis). Kennedy and Steele (1971) attributed similar ingestion of hydrozoans and algae as incidental to the consumption of sought-after clinging and encrusting fauna including amphipods living in constructed tubes firmly attached to ectoproct stalks.

Generally, feeding stanzas of winter flounder of Sable Island Bank changed from exploitation of small, exposed infauna and epifauna by younger fish to consumption of larger epifauna by mature fish. Moreover, the forage of winter flounder changed from the sediment surface to ectoproct colonies as the fish grew. Olla *et al.* (1969) and Keats (1990) both describe P. americanus as a visual predator that characteristically feeds during daylight hours.

There is a tendency for fish to become discriminant feeders and, because of structural or behavioral differences among species, segregate food resources efficiently (Tyler 1972, McEachran *et al.* 1976, Ringler 1979, Wakabara *et al.* 1982). At broad taxonomic levels (Table 2), the three flatfish species, herein, appeared to feed upon similar prey, but as other diet studies of these or related fish species have shown, little or no overlap exists with respect to principal prey consumption (Skalkin 1959, Tyler 1972, Kislalioglu and Gibson 1977, Hacunda 1981, Wakabara *et al.* 1982, Martell 1992).

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BIBLIOGRAPHY

- Appy, T.A., L.E. Linkletter and M.J. Dadswell. 1980. A guide to the marine flora and fauna of the Bay of Fundy: Annelida: Polychaeta. Fish. Mar. Serv. Tech. Rep. No. 920: 124 pp.
- Barnard, J.L. 1969. The Families and Genera of Marine Gammaridean Amphipoda. United States National Museum Bull. No. 271: 535 pp.
- Barnes, R.D. 1974. Invertebrate Zoology. W.B. Sanders Co., Toronto. 870 pp.
- Biernbaum, C.K. 1979. Influence of sedimentary factors on the distribution of benthic amphipods of Fishers Island Sound, Connecticut. J. exp. mar. Biol. Ecol. 38: 201-223.
- Bousfield, E.L. 1973. Shallow Water Gammaridean Amphipoda of New England. Cornell University Press, Ithaca, N.Y. 312 pp.
- Bowman, T.E. 1973. Pelagic Amphipods of the Genus Hyperia and closely related genera (Hyperidea: Hyperiididae). Smithsonian Contributions to Zoology. No. 136: 76 pp.
- Collie, J.S. 1987. Food selection by yellowtail flounder (Limanda ferruginea) on Georges Bank. Can. J. Fish. Aquat. Sci. 44: 357-367.
- Coull, B.C. and S.S. Bell. 1983. Biotic assemblages and communities. In: Bliss, D.E. (ed.) The Biology of Crustacea: Volume 7. Behavior and Ecology.
- Crow, M.E. 1978. Multivariate statistical analysis of stomach contents. In: Lipovsky, S.J. and C.A. Simenstad (eds). Fish Food Habits Studies: Proceedings of the Second Pacific Northwest Technical Workshop. Washington Sea Grant Publication, University of Washington, Seattle. pp. 87-96.
- Dawson, A.S. and A.S. Grimm. 1980. Quantitative seasonal changes in the

- protein, lipid and energy content of the carcass, ovaries and liver of adult female plaice, Pleuronectes platessa L. J. Fish. Biol. 16: 493-504.
- Delbeek, J.C. and D.D. Williams. 1988. Feeding selectivity of four species of sympatric stickleback in brackish-water habitats in eastern Canada. J. Fish Biol. 32: 41-62.
- Fader, G.B., L.H. King, and B. MacLean. 1977. Surficial geology of the eastern Gulf of Maine and Bay of Fundy. Mar. Sci. Pap. 19: 23 p + chart.
- Fauchald, K. and P.A. Jumars. 1979. The diet of worms: A study of polychaete feeding guilds. Oceanogr. Mar. Biol. Ann. Rev. 17: 193-284.
- Gibson, R.N. and I.A. Ezzi. 1987. Feeding relationships of a demersal fish assemblage on the west coast of Scotland. J. Fish Biol. 31: 55-69.
- Gosner, K.L. 1971. Guide to Identification of Marine and Estuarine Invertebrates. Wiley and Sons, Inc., Toronto. 693 pp.
- Grosslein, M.D., R.W. Langton, and M.P. Sissenwine. 1980. Recent fluctuations in pelagic fish stocks of the Northwest Atlantic, Georges Bank region in relation to species interactions. Rapp. P.-v. Réun. Cons. int. Explor. Mer. 177: 374-404.
- Hacunda, J.S. 1981. Trophic relationships among demersal fishes in a coastal area of the Gulf of Maine. Fish. Bull. 79: 775-788.
- Hahm, W. and R. Langton. 1984. Prey selection based on predator/prey weight ratios for some northwest Atlantic fish. Mar. Ecol. Prog. Ser. 19: 1-5.
- Holdich, D.M. and J.A. Jones. 1983. Tanaids: Keys and Notes for the Identification of the Species. Cambridge University Press, New York. 98 pp.
- Jones, N.S. 1952. The bottom fauna and the food of flatfish off the Cumberland coast. J. Anim. Ecol. 21: 182-205.
- Jones, N.S. 1976. British Cumaceans. Academic Press, New York. 61 pp.
- Keats, D.W. 1990. Food of winter flounder Pseudopleuronectes americanus in a sea urchin dominated community in eastern Newfoundland. Mar. Ecol. Prog. Ser. 60: 13-22.
- Kennedy, V.S. and D.H. Steele. 1971. The winter flounder (Pseudopleuronectes americanus) in Long Pond, Conception Bay, Newfoundland. J. Fish. Res. Bd. Canada. 28: 1153-1165.
- King, L.H. 1970. Surficial geology of the Halifax-Sable Island map area. Mar. Sci. Pap. 1: 16 + chart.
- Kislalioglu, K. and R.N. Gibson. 1977. The feeding relationship of shallow water fishes in a Scottish sea loch. J. Fish Biol. 11: 257-266.
- Langton, R.W. 1983. Food habits of yellowtail flounder, Limanda ferruginea (Storer), from off the northeastern United States. Fish. Bull. 81: 15-22.
- Langton, R.W. and R.E. Bowman. 1981. Food of eight northwest Atlantic pleuronectiform fishes. U.S. Dept. Commer., NOAA Tech. Rep. NMFS SSRF-749. 16 pp.
- Levings, C.D. 1974. Seasonal changes in feeding and particle selection by winter flounder (Pseudopleuronectes americanus). Trans. Am. Fish. Soc. 4: 828-832.
- Lincoln, R.J. 1979. British Marine Amphipoda; Gammaridea. British Museum (Natural History), London. 658 pp.
- Libey, G.S. and C.F. Cole. 1979. Food habits of yellowtail flounder, Limanda ferruginea (Storer). J. Fish. Biol. 15: 371-374.
- Macdonald, J.S. and R.H. Green. 1983. Redundancy of variables used to describe importance of prey species in fish diets. Can. J. Fish. Aquat. Sci. 40: 635-637.
- Macdonald, J.S. and R.H. Green. 1986. Food resource utilization by five species of benthic feeding fish in Passamaquoddy Bay, New Brunswick. Can. J. Fish. Aquat. Sci. 43: 1534-1546.
- MacKinnon, J.C. 1972. Summer storage of energy and its use for winter metabolism and gonad maturation in American plaice (Hippoglossoides platessoides). J. Fish. Res. Bd. Canada. 29: 1749-1759.
- Martell, D.J. 1992. Diets of sympatric Pleuronectids (Hippoglossoides

- platessoides, Limanda ferruginea, Pseudopleuronectes americanus) from Sable Island Bank and implications for transmission of sealworm, Pseudoterranova decipiens (Nematoda: Ascaridoidea). M.Sc. Thesis. Dalhousie University, Halifax, N.S., Canada.
- McClelland, G., R.K. Misra, and D.J. Marcogliese. 1983. Variations in abundance of larval anisakines, sealworm (Phocanema decipiens) and related species in cod and flatfish from the southern Gulf of St. Lawrence (4T) and the Breton Shelf (4Vn). Can. Tech. Rep. Fish. Aquat. Sci. No. 1201, ix+51 pp.
- McClelland, G., R.K. Misra, and D.J. Martell. 1990. Larval anisakine nematodes in various fish species from Sable Island Bank and vicinity. Can. Bull. Fish. Aquat. Sci. 22: 83-118.
- McEachran, J.D., D.F. Boesch and J.A. Musick. 1976. Food division within two sympatric species-pairs of skates (Pisces: Rajidae). Mar. Biol. 35 301-317.
- McLeese, J.M. and T.W. Moon. 1989. Seasonal changes in the intestinal mucosa of winter flounder, Pseudopleuronectes americanus (Walbaum), from Passamaquoddy Bay, New Brunswick. J. Fish Biol. 35: 381-393.
- Minet, J.P. 1973 Food and feeding of the American plaice (Hippoglossoides platessoides) on Saint-Pierre Bank (ICNAF Subdiv. 3Ps) and on Cape Breton Shelf (ICNAF Subdiv. 4Vn). ICNAF Res. Doc. 73/34. 19 pp.
- Moore, J.W. and I.A. Moore. 1976. The basis of food selection in flounders, Platichthys flesus (L.), in the Severn Estuary. J. Fish Biol. 9: 139-156.
- Olla, B.L., R. Wicklund and S. Wilk. 1869. Behavior of winter flounder in a natural habitat. Trans. Am. Fish. Soc. 4: 717-720.
- Pitt, T.K. 1973. Food of American Plaice (Hippoglossoides platessoides) from the Grand Bank, Newfoundland. J. Fish. Res. Bd. Canada. 30: 1261-1273.
- Powles, P.M. 1965. Life history and ecology of American plaice (Hippoglossoides platessoides F.) in the Magdalen Shallows. J. Fish. Res. Bd. Canada. 22: 565-598.
- Ringler, N.H. 1979. Prey selection by benthic fishes. In: Clepper, H. (ed.). Predator-prey Systems in Fisheries Management. Washington, Sport Fishing Institute. pp. 219-229.
- Sainte-Marie, B and P. Brunel. 1985. Suprabenthic gradients of swimming activity by cold-water gammaridean amphipod Crustacea over a muddy shelf in the Gulf of St. Lawrence. Mar. Ecol. Prog. Ser. 23: 57-69.
- Schneppenheim, R. and R. Weigmann-Haass. 1986. Morphological and electrophoretic studies of the genus Themisto (Amphipoda: Hyperiidea) from the South and North Atlantic. Polar Biol. 6: 215-225.
- Schoener, T.W. 1971. Theory of feeding strategies. In: Johnston, R.F., P.W. Frank and C.D. Michener (eds). Annual Review of Ecology and Systematics. Annual Reviews Inc., Palo Alto. pp. 369-403.
- Scott, J.S. 1975a. Incidence of trematode parasites of American plaice (Hippoglossoides platessoides) of the Scotian Shelf and Gulf of St. Lawrence in relation to fish length and food. J. Fish. Res. Board Can. 32: 479-483.
- Scott, J.S. 1975b. Intestinal trematode parasites (Trematoda: Digenea) and food of yellowtail flounder (Limanda ferruginea (Storer 1839)) from the Scotian Shelf and Gulf of St. Lawrence. Fish. Mar. Serv. Res. Dev. Tech. Rep. 584: 12 p.
- Scott, J.S. 1976. Digenetic trematode parasites and food of winter flounder (Pseudopleuronectes americanus (Walbaum, 1792)) from the Scotian Shelf and Gulf of St. Lawrence. Fish. Mar. Serv. Res. Dev. Tech. Rep. 618: 9 p.
- Scott, W.B. and M.G. Scott. 1988. Atlantic Fishes of Canada. Can. Bull. Fish. Aquat. Sci. 219: 731 p.
- Shultz, G.A. 1969. The Marine Isopod Crustaceans. Wm. C. Brown Co. Publishers, Dubuque, Iowa. 359 pp.
- Skalkin, V.A. 1959. Food and food interrelationships of flounders of the Ill'insk Bank. Fish. Res. Bd. Canada Trans.

Series No. 789. 35 pp.

- Smith, R.I. (ed.) 1964. Keys to the Marine Invertebrates of the Woods Hole Region, Woods Hole, Mass. Woods Hole Marine Biological Laboratory. 208 pp.
- Squires, H.J. 1990. Decapod Crustacea of the Atlantic Coast of Canada. Can. Bull. Fish. Aquat. Sci. 221: 532 pp.
- Stoner, A.W. 1979. Species specific predation on amphipod crustacea by the pinfish Lagodon rhomboides: mediation by macrophyte standing crop. Mar. Biol. 55: 201-207.
- Tyler, A.V. 1972. Food resource division among northern, marine, demersal fishes. J. Fish. Res. Bd. Canada. 29: 997-1003.
- Wakabara, Y., E. Kawakami de Rezende, and A.S. Tararam. 1982. Amphipods as one of the main food components of three pleuronectiforms from the continental shelf of south Brazil and north Uruguay. Mar. Biol. 68: 67-70.
- Wildish, D.J., A.J. Wilson, and B. Frost. 1989. Benthic macrofaunal production of Browns Bank, Northwest Atlantic. Can. J. Fish. Aquat. Sci. 46: 584-590.
- Wildish, D.J., A.J. Wilson, and B. Frost. 1992. Benthic boundary layer macrofauna of Browns Bank, Northwest Atlantic, as potential prey of juvenile benthic fish. Can. J. Fish. Aquat. Sci. 49: 91-98.
- Worobec, M.N. 1984. Field estimates of the daily ration of winter flounder, Pseudopleuronectes americanus (Walbaum), in southern New England salt pond. J. Exp. Mar. Biol. Ecol. 77: 183-196.

Table 1. Sample sizes of flatfish collections : Numbers of American plaice (*Hippoglossoides platessoides*), yellowtail flounder (*Pleuronectes ferrugineus*) and winter flounder (*Pleuronectes americanus*) stomachs which contained food (N_f) and total number of stomachs (N_t) collected from Sable Island Bank in February and June 1989.

Length groups (cm)	February						June					
	American plaice		yellowtail flounder		winter flounder		American plaice		yellowtail flounder		winter flounder	
	N _f	N _t	N _f	N _t	N _f	N _t	N _f	N _t	N _f	N _t	N _f	N _t
≤15	4	20	19	23	0	0	10	12	0	0	0	0
16-20	15	33	16	24	0	0	23	23	14	16	3	4
21-25	11	33	14	32	0	9	30	31	30	30	30	30
26-30	9	37	13	31	0	14	27	31	30	31	30	30
31-35	16	32	12	30	0	8	24	30	30	32	27	29
36-40	7	18	7	24	0	4	15	23	27	30	22	24
≥41	3	10	1	3	0	2	3	4	0	0	4	4
Totals	65	183	82	167	0	37	132	154	131	139	116	121

Table 2. Frequency of occurrence of all prey species consumed by American plaice (*Hippoglossoides platessoides*), yellowtail flounder (*Pleuronectes ferrugineus*), and winter flounder (*Pleuronectes americanus*) from all length groups and both sampling seasons collected from Sable Island Bank during February and June 1989. Occurrences based on proportion of stomachs that contained food in which a prey species occurred. Levels are represented as <5% ('+'), 5-49% ('++'), and >49% ('+++').

Prey taxa	American plaice	yellowtail flounder	winter flounder
CNIDERIA			
Anthozoa			
<u>Octocorallia</u>			
Alcyonacea			
unk. sp.		+	+
<u>Zoantharia</u>			
Zoanthidea			
<i>Epizoanthus incrustatus</i>			+
Actiniaria			
Athenaria			
unk. sp.			+
Thenaria			
unk. sp.	+	+	+
<i>Actinauge longicornis</i>	++	+	++
RHYNCHOCOELA			
unk. sp.		+	+
TARDIGRADA			
Heterotardigrada			
unk. sp.		+	
ECTOPROCTA			
unk. spp.	+	+	++
MOLLUSCA			
Gastropoda			
unk. sp. (foot)	+	+	+
<u>Prosobranchia</u>			
Archaeogastropoda			
<i>Margarites costalis</i>	+	+	
Mesogastropoda			
<i>Polinices</i> sp.		+	+
Neogastropoda			
<i>Buccinium undatum</i>	+		
<i>Neptunea</i> sp.	+	+	
<u>Opisthobranchia</u>			
Cephalaspidea			
<i>Retusa obtusa</i>	+	+	++
<i>Cylichna gouldii</i>		+	
Bivalvia			
<i>Arctica islandica</i>	+	++	+
<u>Prionodesmata</u>			
Protobranchia			
<i>Nucula delphinodata</i>		+	
<i>Yoldia sapotilla</i>		+	+
<u>Pteriomorphia</u>			
Ptereoconchida			
<i>Musculus discors</i>		+	+
<i>Chlamys islandicus</i>	+		
<i>Anomia</i> sp.	+		

Table 2. (cont'd)

Prey taxa	American plaice	yellowtail flounder	winter flounder
Teleodesmata			
Heterodontida			
<i>Cerasotoderma pinnulatum</i>	+		
<i>Clinocardium ciliatum</i>	+	+	+
<i>Petricola pholadiformis</i>	+	+	
<i>Abra</i> sp.		+	
<i>Solen viridis</i>		+	
<i>Hiatella</i> sp.		+	
<i>Ctyodaria siliqua</i>		+	
ANNELIDA			
Polychaeta			
Phyllodoce			
Phyllodoceidae			
unk. sp.		+	+
<i>Eteone flava</i>			+
<i>Eteone heteropoda</i>		+	+
<i>Eteone longa</i>		+	++
<i>Eteone trilineata</i>		+	
<i>Eteone</i> sp.		+	++
<i>Phyllodoce</i> sp.		++	++
<i>Phyllodoce maculata</i>			+
<i>Phyllodoce mucosa</i>	+	+	++
Polynoidae			
unk. sp.	+	++	++
<i>Enipo</i> sp.			+
<i>Harmothoe extenuata</i>			+
<i>Harmothoe imbricata</i>			+
<i>Harmothoe</i> sp.	+	+	++
<i>Hartmania</i> sp.		+	+
Sigalionidae			
<i>Leanira</i> (nr) <i>hystrices</i>		+	
<i>Leanira</i> sp.		+	
<i>Neoleanira</i> sp.		+	
<i>Pholoe minuta</i>		+	+
Glyceridae			
unk. sp.		+	
<i>Glycera capitata</i>		+	+
Goniadidae			
<i>Gonaida maculata</i>		++	+
<i>Gonaida</i> sp.		+	+
Nephtyidae			
<i>Aglaophamus malmgreni</i>	+	+	
<i>Nephtys bucera</i>		+	
<i>Nephtys caeca</i>			+
<i>Nephtys neotena</i>	+	+	+
<i>Nephtys</i> sp.	++	++	++
Syllidae			
unk. sp.		+	+
<i>Autolytus</i> sp.		+	+
<i>Brania</i> sp.		+	+
<i>Exogone hebes</i>	+	++	+
<i>Streptosyllis arenae</i>		+	

Table 2. (cont'd)

Prey taxa	American plaice	yellowtail flounder	winter flounder
Nereidae			
<i>Nereis pelagica</i>			+
<i>Nereis zonata</i>		+	
Capitellida			
Scalibregmidae			
<i>Scalibregma</i> sp.		+	
Maldanidae			
unk. sp.	+	++	++
<i>Euclymene zonalis</i>		+	+
<i>Euclymene</i> sp.		+	+
<i>Maldanopsis elongata</i>		+	
<i>Nichomache lumbricalis</i>		+	+
<i>Petaloproctus</i> sp.			+
<i>Praxillella patemissa</i>		+	
<i>Praxillella</i> sp.		+	+
<i>Praxillura</i> sp.	+	+	+
Opheliidae			
<i>Ophelia bicornis</i>		+	
<i>Ophelia limacina</i>	+	+	+
<i>Ophelia</i> sp.		+	
<i>Ophelina acuminata</i>	+	++	+
<i>O. cylindricaudata</i>			+
<i>Ophelina</i> sp.		+	
<i>Travisia forbesi</i>			+
<i>Travisia</i> sp.		+	
Spionida			
Spionidae			
unk. sp.	+	++	++
<i>Polydora</i> sp.	+	++	++
<i>Prionospio steenstrupi</i>		+	+
<i>Prionospio</i> sp.		+	+
<i>Scolecopsis squamata</i>		+	
<i>Spio</i> sp.		+	+
<i>Spiophanes bombyx</i>		++	+
Paraonidae			
unk. sp.			+
<i>Aricidea suecica</i>	+	++	
<i>Aricidea wassi</i>		+	
<i>Aricidea</i> sp.	+	+	+
<i>Paraonis</i> sp.		+	
Eunicida			
Onuphidae			
<i>Onuphis conchylega</i>	+		
Lumbrinereidae			
<i>Lumbriclymene</i> sp.		+	
<i>Lumbrineris acuta</i>		+	
Dorvilleidae			
unk. sp.		+	
Aricida			
Orbiniidae			
<i>Scoloplos armiger</i>		++	

Table 2. (cont'd)

Prey taxa	American plaice	yellowtail flounder	winter flounder
<u>Cirratulida</u>			
Cirratulidae			
unk. sp.	+	+	+
<i>Cirratulus cirratus</i>		+	++
<i>Chaetozone setosa</i>		++	+
<i>Chaetozone</i> sp.			+
<i>Tharyx</i> sp.		+	+
<u>Oweniida</u>			
Oweniidae			
<i>Owenia fusiformis</i>		+	+
<u>Terebellida</u>			
Pectinariidae			
<i>Pectinaria granulata</i>	+	++	++
Ampharetidae			
unk. sp.	+	+	+
<i>Ampharete</i> sp.	+	++	++
<i>Ampharete acutifrons</i>		+	
<i>Ampharete baltica</i>		+	
<i>Ampharete lindstroemi</i>	+	++	++
<i>Asabellides</i> sp.		+	
<i>Sabellides</i> (nr) <i>borealis</i>	+	+	+
<i>Sabellides</i> sp.			+
<i>Samytha sexcirrata</i>		+	
Terebellidae			
unk. sp.	+		++
<i>Neoamphitrite</i> sp.			+
<i>Polycirrus eximius</i>		+	
<i>Thelepus cincinnatus</i>			+
<i>Trichobranchus glacialis</i>	+		
<u>Flabelligerida</u>			
Flabelligeridae			
<i>Brada villosa</i>			+
<i>Pherusa affinis</i>	+	+	
<i>Perusa plumosa</i>		+	+
<u>Sabellida</u>			
Sabellidae			
unk. sp.		+	+
<i>Chone dunei</i>		+	
<i>Chone</i> sp.	+	+	+
<i>Euchone incolor</i>	+	++	+
<i>Euchone</i> sp.	+	+	
<i>Potamilla reniformis</i>	+	+	++
<i>Potamilla</i> sp.			+
Serpulidae			
<i>Spirorbis</i> sp.		+	
Oligochaeta			
unk. sp. (marine)		+	
SIPUNCULIDA			
unk. sp.		+	+

Table 2. (cont'd)

Prey taxa	American plaice	yellowtail flounder	winter flounder
ARTHROPODA			
Crustacea			
Copepoda			
unk. parasitic sp.	+		
Calanoida			
unk. sp.	++	+	+
Cyclopoida			
unk. sp.	+		+
Harpacticoida			
unk. sp.		+	+
Brachyura			
Cancridea			
Cancridae			
<i>Cancer borealis</i>		+	+
Malacostraca			
Peracarida			
Cumacea			
Bodotriidae			
<i>Leptocuma minor</i>	+	+++	++
Leuconidae			
<i>Eudorellopsis deformis</i>	+	++	
Lampropidae			
<i>Lamprops quadrireplicata</i>	+	++	++
Diastylidae			
<i>Diastylis sculpta</i>	+	+	+
<i>Diastylis quadrispinosa</i>	++	++	++
Pseudocumidae			
<i>Petalosarsia declivis</i>	+	+	
Tanaidacea			
Paratanaidae			
<i>Leptognathia breviremis</i>		++	+
<i>Leptognathia caeca</i>		+	
Isopoda			
Cryptoniscidae			
unk. sp.		+	+
Flabellifera			
Limnoridae			
<i>Limnoria lignorum</i>		+	
Valvifera			
Idoteidae			
<i>Chiridotea</i> sp.	++	++	
<i>Idotea phosporea</i>	+	+	
<i>Edotea montosa</i>	+	+	++
Asellota			
Janiridae			
<i>Janira alta</i>	+		
Amphipoda			
Hyperidea			
Hyperiididae			
<i>Themisto compressa</i>	++	+	
Gammaridea			
Ampeliscidae			
<i>Ampelisca macrocephala</i>	+	++	++

Table 2. (cont'd)

Prey taxa	American plaice	yellowtail flounder	winter flounder
Ampithoidae			
<i>Ampithoe rubricata</i>		+	
<i>Ampithoe valida</i>		+	
<i>Ampithoe</i> sp.		+	
Aoridae			
<i>Unciola irrorata</i>	++	++	++
Argissidae			
<i>Argissa hamatipes</i>	+	+	++
Corophiidae			
<i>Corophium bonelli</i>	+		
<i>Corophium crassicorne</i>	++	++	++
<i>Corophium volutator</i>		+	
<i>Erichthonius rubricornis</i>	++	++	++
<i>Erichthonius braziliensis</i>			+
Eusiridae			
<i>Rhachotropis lobata</i>	+	+	
<i>Rhachotropis oculata</i>	+		
Gammaridae			
<i>Elasmopus laevis</i>		+	
<i>Elasmopus pocillimanus</i>		+	
<i>Gammarus</i> sp.		+	
<i>Melita dentata</i>		+	
Haustoriidae			
<i>Amphiporeia lawrenciana</i>		+	
<i>Priscillina armata</i>	+	++	+
<i>Bathyporeia quoddyensis</i>		++	
Ischyroceridae			
<i>Ischyroceros anguipes</i>	++	++	++
Leucothoidae			
<i>Leucothoe spinicarpa</i>			+
Lysianassidae			
<i>Anonyx lilljeborgii</i>	+	+	
<i>Anonyx sarsi</i>		+	
<i>Hippomedon serratus</i>	+	+	+
<i>Orchomenella minuta</i>		+	
<i>Orchomenella pinguis</i>		+	+
<i>Tryphosella</i> sp.		+	
Oedicerotidae			
<i>Monoculodes edwardsi</i>	++	++	++
<i>Monoculodes intermedius</i>	+	++	+
Photidae			
<i>Leptocheirus pinguis</i>	+	++	++
<i>Leptocheirus plumosus</i>		+	+
<i>Podoceropsis nitida</i>		+	+
<i>Photis</i> sp.	++	+++	++
<i>Protomedeia fasciata</i>	+	+	++
Phoxocephalidae			
<i>Phoxocephalus holbolli</i>		+	
<i>Paraphoxus spinosus</i>			+
<i>Harpinia propinqua</i>		+	
Pleustidae			
<i>Pleustes panopla</i>	+		+
<i>Pleusymtes glaber</i>	+		+
<i>Stenopleustes gracilis</i>			+

Table 2. (cont'd)

Prey taxa	American plaice	yellowtail flounder	winter flounder
Pleustidae (cont'd)			
<i>Stenopleustes inermis</i>	+	+	++
Podoceridae			
<i>Dyopedos</i> sp.	+	++	++
Pontogeneiidae			
<i>Pontogeneia inermis</i>	+	++	++
Stenothoidae			
<i>Metopa alderi</i>	+	+	+
<i>Metopa borealis</i>		+	+
<i>Metopa bruzelii</i>	+	++	++
<i>Metopa propinqua</i>		+	+
<i>Metopa pusilla</i>	+	+	+
<i>Metopella augusta</i>	+	+	+
<i>Metopelloides</i> sp.		+	
<i>Parametopella cypris</i>		+	+
<i>Stenothoe brevicornis</i>			
var. <i>canadensis</i>	+		+
<i>Stenothoe gallensis</i>			+
<i>Stenothoe minuta</i>			+
Caprellidea			
Caprellidae			
<i>Aeginina longicornis</i>	++		++
Mysidacea			
Mysida			
<i>Erythroops erythroptalma</i>	++	+	+
<i>Mysis mixta</i>	++	+	
<i>Neomysis americana</i>	+		
Decapoda			
Caridea			
Hippolytidae			
<i>Spirontocaris lilljeborgii</i>	+		
Pandalidae			
<i>Dichelopandalus leptocerus</i>	+	+	
<i>Pandalus borealis</i>	+	+	
Crangonidae			
<i>Crangon septemspinosa</i>	++	+	+
Paguroidea			
Paguridae			
unk. sp. (glaucothoe)	+		+
unk. sp. (late stage larva)	+	+	
<i>Pagurus</i> sp.	++	+	++
ECHINODERMATA			
Holothuroidea			
unk. sp.		+	
Dendrochirotida			
Cucumariidae unk. sp.			+
Molpadiida unk. sp.		+	+
Echinoidea			
Echinoida			
<i>Strongylocentrotus droebachiensis</i>	+	++	+
<i>Echinarachnius parma</i>	+	++	+

Table 2. (cont'd)

Prey taxa	American plaice	yellowtail flounder	winter flounder
<u>Clypeasteroidea</u>			
<i>Echinarachnius parma</i>	+	++	+
Stellaroidea			
<u>Ophiuroidea</u>			
<i>Ophiura sarsi</i>		+	
<i>Amphipholis squamata</i>	++	++	+
CHORDATA			
Ascidacea			
<u>Pleurogona</u>			
<i>Molgula sp.</i>	+	+++	+++
Osteichthyes			
<u>Perciformes</u>			
Stichaeidae			
unk. sp.		+	
Ammodytidae			
<i>Ammodytes sp.</i>	+	++	+

Table 3. Occurrence (O) of general prey taxa consumed by *Hippoglossoides platessoides* (Hp), and *Pleuronectes ferrugineus* (Pf) collected from Sable Island Bank in February 1989. Within each length group, occurrence is calculated as the proportion (percent) of stomachs that contain food in which a prey taxa was found.

Prey group	Predator length groups (cm)													
	≤15		16-20		21-25		26-30		31-35		36-40		≥41	
	Hp	Pf	Hp	Pf	Hp	Pf	Hp	Pf	Hp	Pf	Hp	Pf	Hp	Pf
Anthozoa	0	0	0	0	18	0	22	0	38	8	0	0	33	0
Ectoprocta	0	0	0	6	0	0	0	8	0	0	0	14	0	0
Mollusca														
Gastropoda	0	0	0	0	0	7	33	15	0	0	0	0	0	100
Bivalva	0	0	0	6	9	14	11	0	13	17	0	0	67	0
Polychaeta														
Phyllodocida	0	42	0	25	9	14	11	39	0	58	0	57	0	0
Capitellida	0	0	0	13	0	14	0	15	0	0	0	29	0	0
Spionida	0	21	0	6	0	7	0	23	0	42	0	43	0	0
Eunicida	0	0	0	0	0	0	0	0	6	0	0	0	0	0
Aricida	0	0	0	0	0	7	0	0	0	8	0	0	0	0
Cirratulida	0	5	0	6	0	14	0	23	0	17	0	29	0	0
Oweniida	0	0	0	0	0	0	0	0	0	0	0	0	0	0
Terebellida	0	37	7	25	0	14	11	46	13	25	0	0	0	0
Flabelligerida	0	0	0	0	0	0	0	31	6	8	0	0	0	0
Sabellida	0	5	0	6	0	0	0	8	6	0	0	0	0	0
Crustacea														
Copepoda	25	11	0	0	0	0	11	8	13	0	0	14	0	0
Brachyura	0	0	0	0	0	7	0	0	0	8	0	0	0	0
Cumacea	0	84	0	56	9	57	0	85	0	92	0	57	0	0
Tanaidacea	0	32	0	6	0	0	0	0	0	8	0	0	0	0
Isopoda	0	11	0	6	0	21	0	31	0	0	14	0	0	0
Amphipoda														
Hyperiidea	0	0	0	0	0	0	0	0	0	0	0	0	0	0
Gammaridea	75	95	33	88	55	93	44	92	50	100	0	100	0	0
Caprellidea	0	0	0	0	0	7	0	0	6	8	14	0	0	0
Mysidacea	75	0	73	0	91	0	11	0	31	0	14	43	0	0
Caridea	0	0	0	0	18	7	11	0	25	0	43	14	0	0
Paguroidea	0	0	0	0	0	0	22	0	13	0	0	0	33	0
Echinodermata														
Holothuroidea	0	0	0	0	0	0	0	8	0	0	0	0	0	0
Echinoidea	0	0	0	6	0	21	0	69	13	58	43	0	0	0
Stellaroidea	0	0	0	0	0	7	11	0	13	0	0	14	0	0
Chordata														
Ascidacea	0	0	0	6	0	7	11	15	0	0	0	0	33	0
Osteichthyes	0	5	0	0	0	0	0	0	0	0	0	0	0	0
Other	0	0	0	0	0	0	0	0	0	8	0	14	0	0

Table 4. Occurrence (O) of general prey taxa consumed by *Hippoglossoides platessoides* (Hp), *Pleuronectes ferrugineus* (Pf), and *Pleuronectes americanus* (Pa) collected from Sable Island Bank in June 1989. Within each length group, occurrence is calculated as the proportion (percent) of stomachs that contain food in which a prey taxa was found.

Prey group	Predator length groups (cm)											
	≤15			16-20			21-25			26-30		
	Hp	Pf	Pa	Hp	Pf	Pa	Hp	Pf	Pa	Hp	Pf	Pa
Anthozoa	0	—	—	0	0	0	0	3	10	11	0	37
Ectoprocta	0	—	—	0	0	0	0	0	7	0	0	17
Mollusca												
Gastropoda	0	—	—	0	14	0	13	7	53	15	14	33
Bivalva	0	—	—	0	7	33	10	17	31	7	3	0
Polychaeta												
Phyllodocida	10	—	—	13	43	0	30	47	40	19	72	50
Capitellida	0	—	—	4	21	0	7	13	27	0	35	33
Spionida	0	—	—	0	29	0	13	33	20	7	35	20
Eunicida	0	—	—	0	7	0	0	7	0	0	0	0
Aricida	0	—	—	0	0	0	0	3	0	0	0	0
Cirratulida	0	—	—	0	14	0	0	33	7	0	59	20
Oweniida	0	—	—	0	0	0	0	0	0	0	0	3
Terebellida	0	—	—	0	64	100	3	67	73	11	76	63
Flabelligerida	0	—	—	0	0	0	0	0	0	0	0	6
Sabellida	0	—	—	0	0	0	3	3	0	11	3	16
Crustacea												
Copepoda	0	—	—	0	0	0	13	3	3	7	3	7
Brachyura	0	—	—	0	0	0	0	0	0	0	0	7
Cumacea	90	—	—	78	86	100	73	90	83	59	90	60
Tanaidacea	0	—	—	0	0	0	0	0	0	0	0	3
Isopoda	10	—	—	13	7	0	7	31	7	11	3	3
Amphipoda												
Hyperideia	30	—	—	48	7	0	40	0	0	22	0	0
Gammaridea	80	—	—	70	86	100	77	83	93	74	79	90
Caprellidea	0	—	—	4	0	0	7	0	13	7	3	27
Mysidacea	30	—	—	26	0	0	30	0	3	30	7	0
Caridea	0	—	—	9	14	0	7	10	3	19	3	3
Paguroidea	0	—	—	4	7	0	0	0	7	4	0	7
Echinodermata												
Holothuroidea	0	—	—	0	7	0	0	0	0	0	0	3
Echinoidea	0	—	—	0	0	0	0	7	3	0	7	13
Stellaroidea	0	—	—	39	0	0	63	3	0	30	0	0
Chordata												
Ascidacea	0	—	—	0	79	100	7	100	83	11	100	57
Osteichthyes	0	—	—	0	7	0	0	17	3	4	14	3
Other	0	—	—	0	7	0	0	7	0	0	31	0

Table 4. (cont'd)

Prey group	Predator length groups (cm)								
	31-35			36-40			≥41		
	Hp	Pf	Pa	Hp	Pf	Pa	Hp	Pf	Pa
Anthozoa	0	0	30	7	7	27	0	—	25
Ectoprocta	4	0	44	7	0	59	0	—	75
Mollusca									
Gastropoda	13	10	4	13	15	14	0	—	0
Bivalva	13	7	11	7	30	0	0	—	0
Polychaeta									
Phyllodocida	4	59	52	7	93	55	0	—	25
Capitellida	0	38	15	0	63	36	0	—	25
Spionida	4	55	30	0	89	27	0	—	25
Eunicida	0	3	0	0	4	0	0	—	0
Aricida	0	3	0	0	33	0	0	—	0
Cirratulida	0	59	19	0	52	23	0	—	0
Oweniida	0	0	0	0	15	0	0	—	0
Terebellida	4	72	56	7	48	68	0	—	100
Flabelligerida	0	0	7	0	0	0	0	—	0
Sabellida	0	3	33	0	56	18	0	—	0
Crustacea									
Copepoda	8	0	0	13	0	5	0	—	0
Brachyura	0	0	4	0	0	0	0	—	0
Cumacea	50	69	33	47	93	59	0	—	50
Tanaidacea	0	10	0	0	4	0	0	—	0
Isopoda	21	10	11	27	22	5	0	—	0
Amphipoda									
Hyperiidea	46	0	0	87	30	0	67	—	0
Gammaridea	58	86	67	40	93	82	33	—	100
Caprellidea	13	0	37	20	15	64	0	—	100
Mysidacea	13	3	0	13	4	0	0	—	0
Caridea	29	3	7	13	4	0	33	—	0
Paguroidea	13	0	15	33	7	5	100	—	0
Echinodermata									
Holothuroidea	0	0	0	0	4	5	0	—	0
Echinoidea	4	17	4	7	37	5	33	—	0
Stellaroidea	42	0	0	7	52	5	0	—	0
Chordata									
Ascidacea	4	97	44	0	41	23	0	—	0
Osteichthyes	0	10	0	0	4	0	0	—	0
Other	0	14	4	0	22	9	0	—	25