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DNA barcoding confirms species substitution of *Parastromateus niger* (black pomfret) using exotic species *Piaractus brachypomus* (Red Bellied Pacu)

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Abstract

Species substitution is a case of fraud practised by the fish dealers for acquiring additional profit. During a fishery survey in the local markets of Kerala state (South India), some ‘alien’ fishes were encountered which were traded along with *Parastromateus niger* as ‘black pomfrets’. Samples of these two groups were collected for morphological and molecular characterization (viz. COI gene). COI sequences of *P. niger* corroborated with the same in NCBI while morphological and molecular characters of ‘alien’ fishes corroborated with an exotic fish *Piaractus brachypomus*. Molecular analysis carried out using additional COI sequences of ‘pomfret’ and ‘piranha’ fishes acquired from NCBI confirmed the speciation of ‘alien fishes’ as *P. brachypomus*. This study reports the illegal trade of *P. brachypomus* as a species substituent for *P. niger* in Kerala and recommends the enforcement of strict laws for preventing food frauds as it could bring up serious economic breakdowns in fisheries sector.

Keywords: *Parastromateus niger*, COI, *Piaractus brachypomus*, species substitution

1. Introduction

Parastromateus niger (Class Actinopterygii, Order Perciformes, Family Carangidae) or black pomfret is a high valued marine fish inhabiting the shallow inshore waters of the Indian Ocean [1, 2]. Increasing demand for this species has resulted in the reduction of its stock to a larger extent due to over exploitation and lack of proper fishing management programs [3]. Recently its landings in Indian coastal areas declined drastically to the tune of 13924 tonnes in 2016 being only 52.36% of its previous year catch [4]. Higher availability of this species is needed to satisfy its increased demand since other sources of supply like aqua farming has not been in vogue for this species. This indicates possibilities of species substitution and misrepresentation using morphologically similar exotic, non-invasive, protected and low valued species, since seafood mislabelling is a global and significant issue in the present scenario [5-10].

In Kerala (S. India), *P. niger* is having higher market acceptance and is traded in fresh, processed (fillets, brined), sun dried and frozen conditions [11]. During a fishery survey conducted in local markets of Kochi (Kerala, S. India), certain ‘alien’ fishes were encountered which were traded along with *Parastromateus niger* (black pomfret) as black pomfrets. *P. niger* was identified on the basis of their morphological characters while the ‘alien’ ones showed a similarity in its body outline and pattern only. Even though, *P. niger* is reported to possess morphological similarity to the white pomfrets, *Pampus argenteus* and Chinese pomfrets, *P. chinensis* [12], little is known about a morphologically similar species substituent for black pomfrets. Hence, whole, fresh samples of these two different groups were collected from local markets of Kochi and a molecular approach was carried out to confirm the case of species misrepresentation. Since mitochondrial cytochrome c oxidase subunit I (COI) gene was used as a well-established molecular marker for identification of bio specimens including invasive and exotic species [13-15], the same were developed from the collected specimens and the results are presented here.

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2 Materials and methods

2.1 Sample collection

Whole fish samples of *Parastromateus niger* (n=40) and ‘alien’ fishes (n=70) were collected from the local markets of Kochi (S. India) and transported to the laboratory in iced condition.

A thorough morphological examination was carried out to identify and confirm the speciation of collected specimens. In order to generate COI sequences, tissue samples were taken from whole fish samples and preserved at -20 °C.

2.2 Genotyping

Genomic DNA was isolated following the spin column protocol for purification of total DNA from animal tissues provided in the DNeasy Blood and Tissue Kit (Qiagen). For amplifying partial mitochondrial COI gene sequences, the primer pair Fish F1 (5'TCAACCAACCACAAAGACATTGGCAC3') and Fish R1 (5'TAGACTTCTGGGTGCCAAAGAATCA 3')^[16] were used. Temperature profile for amplification included 35 repeats of denaturation at 92 °C for 1 min, annealing at 48 °C for 35-50 s and extension at 68 °C for 90-120 sec^[16]. Sigma Aldrich ReadyMix™ Taq with MgCl² was the PCR reaction

mix used for amplifying COI gene sequences in a Corbett gradient thermal cycler. Following the agarose gel electrophoresis using a 1.2% agarose gel, PCR amplicons showing intense bands were chosen for purification and sequencing. Sequences were manually edited, translated online and compiled using BioEdit 7.0.9^[17] followed by alignment using Clustal X^[18]. Phylogenetic tree based on Neighbour Joining (NJ) analysis as well as nucleotide sequence distance data were generated (Kimura 2-Parameter model) using MEGA 5^[19].

3. Results and discussion

The collected species were differentiated morphologically as *Piaractus brachypomus* (Class Actinopterygii, Order Characiformes Family Characidae) following^[20-22] and *Parastromateus niger* following^[23] and the major morphological characters identified are detailed in Table 1.

Table 1. Major morphological characters for differentiating *Parastromateus niger* and *Piaractus brachypomus*

Species	<i>Parastromateus niger</i>	<i>Piaractus brachypomus</i>
Body	Body elevated, compressed and rounded anterior profile	Deep and laterally compressed body
Colour	Deep/greyish brown with blue reflections. Cheeks, opercle and abdomen likely same colour and pale/brownish. Black colour present in pectoral and caudal edge	Deep silvery/grey body with silver shades on sides and dark dorsal portion Red colouration on belly chin and pectoral fins and leading rays of anal fin.
Scales	Small, deciduous, completely covering dorsal and anal fins	The scales are numerous cycloid with smooth caudal margin
Lateral Line	Lateral line very weakly arched anteriorly, with junction of straight and curved parts below posterior third of dorsal fin; straight part of lateral line with 8 to 19 weak scutes. 76 scales in lateral line, 35 of which are in straight portion, and scarcely armed	Lateral line is roughly straight Lateral line scales ranges between 102- 117
Keel	Strong keel present in the belly portion	A serrated keel runs along the belly region and extends upto to anus
Dentition	Teeth in jaws, slender, pointed and inserted on ridge	Two series of molariform incisors located on premaxilla and one row of dentary teeth
Fins	Dorsal – 5 spines, 42-44 rays Pectoral – 1 spine, 22 rays Anal – 3 spines, 33-35 rays Caudal – 19 rays	Dorsal – 2 spines, 15-18 rays Pectoral – 1 spine, 16-19 rays Anal – 3 spines, 24-28 rays Pelvic – 1 spine, 7 rays
Adipose fin	Absent	A small, unrayed adipose fin is present approximately midway between the dorsal and caudal fins with base lined with scales

3.1 Genotyping results

Molecular analysis using the primer pair and temperature profile generated COI gene sequences which were submitted in NCBI (Table 2). These COI sequences were subjected to primary BLAST search^[24], and they showed higher identity (>97%) towards homologous COI sequences as *Piaractus*

brachypomus and *Parastromateus niger* respectively, present in public database. Molecular analyses were carried out using additional acquired homologous COI sequences^[25] of ‘Piranha’ and ‘Pomfret’ fishes from NCBI. The details regarding incorporated COI sequences are detailed in Table 2.

Table 2: Details of COI sequences incorporated for molecular analyses

SI No	TAXON	Number of COI sequences	Accession numbers	Status
1	<i>Piaractus brachypomus</i>	7	MF693914-20	Developed for the present study
		1	KP723356	Acquired from NCBI
2	<i>Parastromateus niger</i>	1	MF737196	Developed for the present study
		3	KU943742 HQ560987 HQ560977	Acquired from NCBI
3	<i>Pampus argenteus</i>	3	FJ226531-33	Acquired from NCBI
4	<i>Pampus chinensis</i>	4	FJ265824-25 FJ226529-30	Acquired from NCBI
5	<i>Brama brama</i>	4	JF492979-82	Acquired from NCBI
6	<i>Brama japonica</i>	3	JF952690 GU440256 FJ164426	Acquired from NCBI
7	<i>Brama orcinii</i>	4	KY371231 KF489508 KM366110 KM252676	Acquired from NCBI
8	<i>Collossoma macropomum</i>	1	HQ420843	Acquired from NCBI
9	<i>Eumegistus illustris</i>	1	KU943868	Acquired from NCBI
10	<i>Taractes asper</i>	2	GU440550	Acquired from NCBI

			EU400170	
11	<i>Taractes rubescens</i>	4	KY372188-91	Acquired from NCBI
12	<i>Taractichthys longipinnis</i>	2	AB639845 EF609476	Acquired from NCBI
13	<i>Taractichthys steindachneri</i>	4	KY372194-96 KY372198	Acquired from NCBI
14	<i>Xenobrama microlepis</i>	3	KX497160-61 EF609495	Acquired from NCBI
15	<i>Poecilia reticulata</i>	1	KR871634	Acquired from NCBI

3.2 Phylogenetic tree and genetic distance data

Phylogenetic tree based on neighbour joining analysis clearly differentiated ‘Pomfret’ and ‘Piranha’ fishes as two different clades. Within the pomfrets fishes, individuals belonging to genus *Brama* (*B. brama*, *B. japonica* and *B. orcinus*), genus *Eumegistus* (*E. illustris*), genus *Xenobrama* (*X. microlepis*), genus *Taractichthys* (*T. longipinnis* and *T. steindachneri*) and genus *Taractes* (*T. asper* and *T. rubescens*) arrayed within a major clade. The neighbouring clade was formed by *Parastromateus niger*, *Pampus argenteus* and *P. chinensis* justifying their relationship with each other. *Pampus argenteus* and *P. chinensis* represented closest relatives of *Parastromateus niger* than the other ‘pomfrets’. COI

sequence developed for *P. niger* in this study also got arrayed with the rest of the same without any confusions. So also in ‘Piranhas’, COI sequences of *Piaractus brachypomus* sequenced for this study and the acquired nucleotide sequence of the same showed genetic congruency. ‘*Collossoma macropomum*’ arrayed next to *P. brachypomus* with high bootstrap value. At intraspecific level, there existed little genetic divergence within *Piaractus brachypomus* and *Parastromateus niger*. However, their higher interspecific divergence made *Piaractus brachypomus* to array among the ‘piranhas’ and *Parastromateus niger* among the ‘pomfrets’ (Fig. 1).

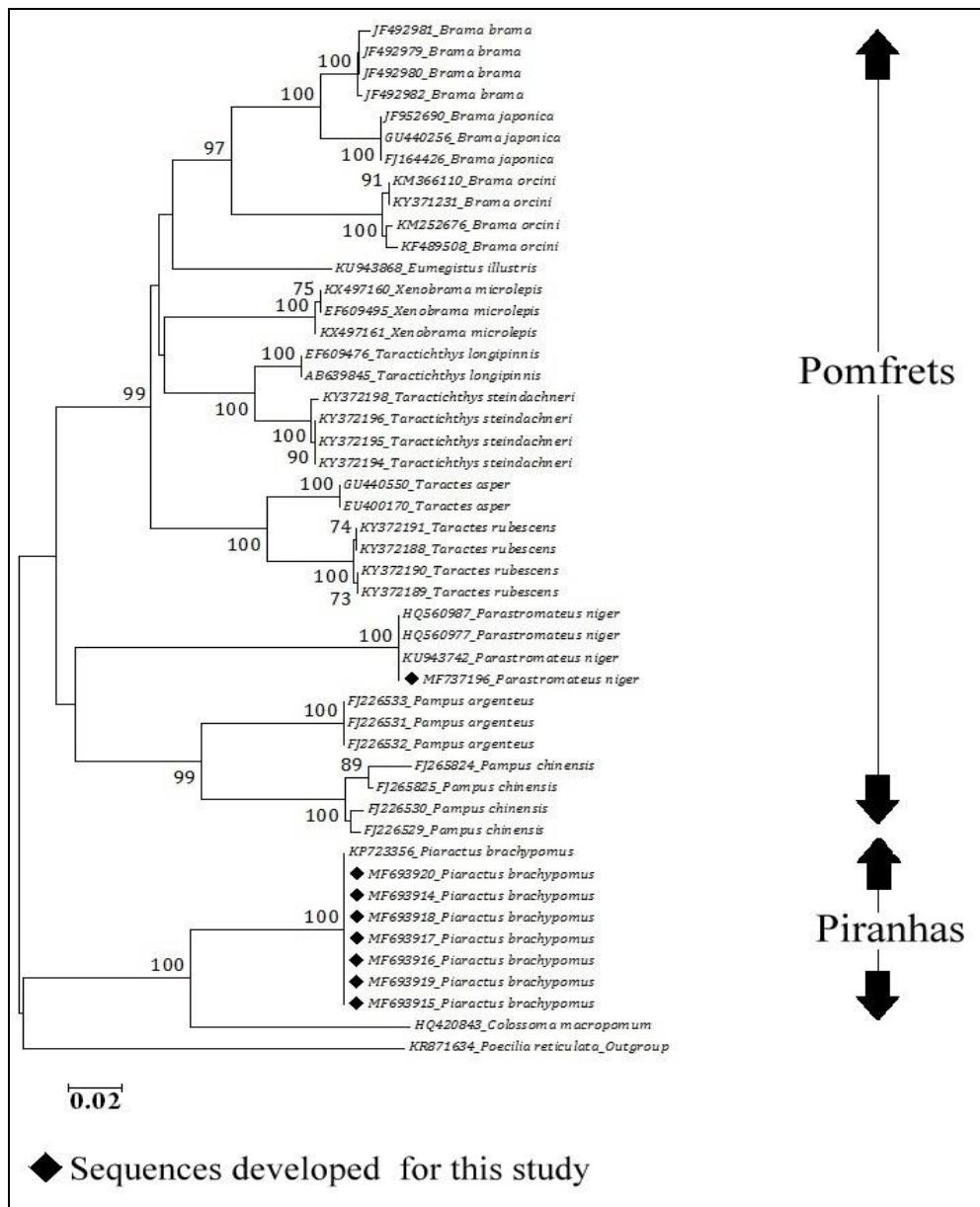


Fig 1: Phylogenetic tree generated using COI sequences with bootstrap values higher than 70

These findings from the phylogenetic tree was further confirmed by pairwise distance data (supplementary material). According to pairwise sequence data, the intraspecific distance within *Piaractus brachypomus* and *Parastromateus niger* was zero. At interspecific level, the divergence rate between *Piaractus brachypomus* and *Parastromateus niger* was very high (25.9%). This justified their differential array (in NJ tree) representing two highly distinct species. Interspecific divergence between other selected individuals (Table 3) also supported the findings of phylogenetic tree.

4. Discussion

Piaractus brachypomus Cuvier, 1818 (Family: Characidae) also known as ‘Red Bellied Pacu’, is the largest scaled fish inhabiting the Amazon and Orinoco basins of South America [26-28]. It is considered as an important food and aquarium fish in many South American countries including Brazil. Moreover, export of *P. brachypomus* populations from its type localities to different countries across the globe has also been reported. In India, *P. brachypomus* was introduced from Bangladesh during 2003-04 [29-31]. Earlier, occurrence of a huge ‘Pomfret’ like silvery fish in the north Western Ghats of India, which was identified as *P. brachypomus* was published [32]. Later, inhabitancy of this species was reported from natural waters of other states within the country including Kerala, Tamil Nadu, Telangana and West Bengal [26, 33, 27, 29, 34, 30, 35, 36]. Hitherto, this species is considered as an unofficially introduced exotic fish that could cause serious problems to humans and native fauna [30, 37, 29, 33].

Aquaculture and breeding of this species gained popularity in Kerala and local fish keepers and fish culturists started culturing the same in ponds and tanks across the state [29]. Recent studies reported its occurrence from various lakes like Vembanad [33] and Vellayani [29] and riverine systems including Pamba [26], Chalakudy [32], Periyar [31] and Muvattupuzha [27] of Kerala. This has been attributed to the accidental release of cultured stocks to open waters [29, 33].

Present work focuses on the use of *Piaractus brachypomus* as a substituent to *Parastromateus niger* in local markets of Kochi, Kerala (S. India) and the mode of fish fraud. Accidental species substitutions occur when two different species exhibiting similar morphological characters obstruct proper identification [38-40]. Multiple species procuring the same vernacular name (and *vice versa*) also creates ambiguities and confusions in species identification [38, 39]. Intentional species substitution is the most common type practised by the dealers for acquiring additional profit. It is a clear case of consumer fraud in which species of high commercial value and high consumer acceptance are intentionally replaced with those having less commercial significance and low market value, leading to economic loss [38, 40-42]. Another case of substitution is the trade of vulnerable

or overexploited species by masking their exploitation rate, affecting their fishing control and management [6, 38, 41-46]. In addition, substitutions may pose health risk to the consumers when economically important species are replaced by poisonous individuals that can cause serious health hazards [41, 42, 47, 48]. Here, *Parastromateus niger* and *Piaractus brachypomus* were traded in local markets as Black pomfrets. Based on thorough morphological examination, these species could be differentiated. However, consumers lacking knowledge regarding the introduction of this exotic fish could fail to identify the same by morphology. Misrepresentation of *Parastromateus niger* having higher market acceptance with a low valued exotic species like *Piaractus brachypomus* seems like an intentional type of species substitution since there will be considerable increase in profit for the dealer/seller and loss for the customer. A similar case of species substitution in whole fish (*Pampus chinensis*) using *Piaractus mesopotamicus* was published by [49]. Progressively, there may be chances for the use of *P. brachypomus* as a species substituent in processed products (like chunks, fillets, dressed frozen etc.) of high valued fishes.

More than 20 provinces of United States have already legally banned pacu and piranha from trade and aquaculture purposes due to their ruinous reaction on natural fish species and ecosystems [32]. They exhibits out-competing nature on the native species for prey and other resources causing ecological disparity. It over powers the local fish species by its huge size and powerful dentition and by feeding on eggs and fry. It is found to disseminate exotic parasites and diseases which may affect the native fauna gravely [26]. There are reports stating the role of *P. brachypomus* in causing outbreaks of parasitic infestations of Klossinemella and Rondonia in Argentina and Brazil, as well as Spectatus infestations in Argentina, Brazil, and Paraguay [50]. However, its potential impacts have been reported and it is causing enough concern to be listed as a potential threat in some states [51]. Their dentition pattern is a threat to humans as it can cause serious bites [52]. Two human deaths caused due to biting off the testicles of fishermen by Pacu was reported from Papua New Guinea [37].

According to [53], almost half of the 12 pacu species are being marked as traumatogenic, which is a behaviour considered dangerous and exhibited in the piranhas. From India also, there are reports revealing its “traumatogenic” effect on humans in Dimbhe reservoir in Maharashtra [54]. Impacts of introduction of Red bellied pacu in Kerala waters are also extensively discussed on the wake of serious damage to the native aquatic species within the state [32] and is feared to cause the disappearance of many economically important indigenous fishery resources from the state. Since *Piaractus brachypomus* is considered as an exotic species, its negative impacts on biodiversity needs to be monitored and standard quarantine procedures need to be enforced [27].

Table 3: Genetic distance data (Intraspecific divergence within *Piaractus brachypomus* and *Parastromateus niger* (Yellow colour) and interspecific divergence between them (Light blue colour))

5. Conclusion

In India, limited quality assessment techniques employed for evaluating the authenticity of locally marketed fish and fishery products are a major problem. Therefore, sensitive and reliable methods should be adopted for the correct identification of traded species, thus preventing species substitutions and market fraud [55]. Present work strongly recommends the need for DNA based studies as a harmonised tool for fishery resource management [42, 56]. Implementation of strict laws for monitoring the release of alien species into natural waters and aquaculture systems are to be done by government according to the scenario persisting within the country/state or else our indigenous fishes and fishery resources will be eradicated by the non-indigenous fishes [36].

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7. Declaration of interest

The authors declare responsibility for the entire contents of this paper and have no conflicts of interest.

8. References

- Tao Y, Mingru C, Jianguo D, Zhenbin L, Shengyun Y. Age and growth changes and population dynamics of the black pomfret (*Parastromateus niger*) and the frigate tuna (*Auxis thazard thazard*), in the Taiwan Strait. Latin American Journal of Aquatic Research. 2012; 40(3):649-656
- Lim KKP, Low JK. A Guide to Common Marine Fishes of Singapore. Singapore Science Centre. 1998, 163
- Sharifi M, Sourinejad I, Hoseyni SJ, Qasemi SA, Faghah A. Genetic diversity of Black Pomfret (*Parastromateus niger*) in Iranian coasts of Persian Gulf using AFLP molecular marker. Journal of Aquatic Ecology. 2012; 2(2):81-68
- CMFRI Annual Report., <http://eprints.cmfri.org.in/> Various Years. 2015, 2016d
- Jacquet JL, Pauly D. The rise of seafood awareness campaigns in an era of collapsing fisheries. Marine Policy. 2007; 31(3):308-313.
- Rasmussen RS, Morrissey MT. DNA-based methods for the identification of commercial fish and seafood species. Comprehensive reviews in food science and food safety. 2008; 7(3):280-295.
- Wong EHK, Hanner RH. DNA barcoding detects market substitution in North American seafood. Food Research International. 2008; 41(8):828-837.
- Jabado RW, Al Ghais SM, Hamza W, Henderson AC, Spaet JL, Shivji MS et al. The trade in sharks and their products in the United Arab Emirates. Biological Conservation. 2015; 181:190-198.
- Khedkar GD, Tiknaik AD, Shinde RN, Kalyankar AD, Ron TB, Haymer D. High rates of substitution of the native catfish Clarias batrachus by *Clarias gariepinus* in India. Mitochondrial DNA Part A. 2016; 27(1):569-574.
- Miller D, Jessel A, Mariani S. Seafood mislabelling: comparisons of two western European case studies assist in defining influencing factors, mechanisms and motives. Fish and fisheries. 2012; 13(3):345-358.
- Joseph MM, Jayaprakash AA. Status of exploited marine fishery resources of India. Central Marine Fisheries Research Institute. Kochi, 2003.
- Tan HH. Observations on the black pomfret, *Parastromateus niger* (Teleostei: Perciformes: Carangidae). Nature in Singapore. 2009; 2:167-169.
- Saunders GW. Routine DNA barcoding of Canadian Gracilariales (Rhodophyta) reveals the invasive species *Gracilaria vermiculophylla* in British Columbia. Molecular ecology resources. 2009; 9(1):140-150.
- Ghahramanzadeh R, Esselink G, Kodde LP, Duistermaat H, Valkenburg JLCH, Marashi SH et al. Efficient distinction of invasive aquatic plant species from non-invasive related species using DNA barcoding. Molecular ecology resources. 2013; 13(1):21-31.
- Yang Li, Tan Z, Wang D, Xue L, Guan MX, Huang T, Li R. Species identification through mitochondrial rRNA genetic analysis. Scientific reports. 2013; 4:4089.
- Escobar LMD, Andrade-López J, Farias IP, Hrbek T. Delimiting evolutionarily significant units of the fish, *Piaractus brachypomus* (Characiformes: Serrasalmidae), from the Orinoco and Amazon river basins with Insight on Routes of Historical Connectivity. Journal of Heredity. 2015; 106(1):428-438
- Hall TA. Bioedit: A user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. Nucleic Acids Symposium series. 1999; 41:95-8.
- Thompson JD, Gibson TJ, Plewniak F, Jeanmougin F, Higgins DG. The Clustal IX windows interface: flexible strategies for multiple sequence alignment aided by quality analysis tools. Nucleic Acids Research. 1997; 24:4876-4882
- Tamura K, Peterson D, Peterson N, Stecher G, Nei M, Kumar S. MEGA5: Molecular Evolutionary Genetics Analysis using Maximum Likelihood, Evolutionary Distance, and Maximum Parsimony Methods. Molecular Biology and Evolution. 2011; 28(10).
- Cuvier. Memoires du museum D'histoire naturelle. Tome quatrieme. 1818t, 452-456
- Hensel K. First record of the pirapitinga *Piaractus brachypomus* (Actinopterygii: Serrasalmidae) in Slovakia. Biologia. 2004; 59(15):205-210.
- Ross ST. The Inland Fishes of Mississippi. Univ. Press of Mississippi. 2001: 624
- Evermann Barton Warren, Seale Alvin. Fishes of the Philippine Islands. Bureau of Fisheries Document No. 1906; 7:65-66.
- Baker CS. A truer measure of the market: the molecular ecology of fisheries and wildlife trade. Molecular Ecology. 2008; 17(18):3985-3998
- Deepak Jose, Harikrishnan M. Non-homologous COI barcode regions: a serious concern in decapod molecular taxonomy. Mitochondrial DNA Part A. 2016; 28(4):482-492.
- Dharan Rani S, Williams Sherly E. First record of the Pirapitinga *Piaractus brachypomus*, Cuvier, 1818 (Actinopterygii: Serrasalmidae) in Pamba river Kerala, India. The Bioscan. 2017; 12(1):121-124.
- Zeena KV, Beevi KJ. On a Report of Red-Bellied Pacu *Piaractus brachypomus* (Cuvier, 1818), (Characiformes: Characidae) from Muvattupuzha River, Kerala, India. Journal of the Bombay Natural History Society (JBNHS). 2014; 111(2):138-140.

28. Jegu M. Serrasalmidae (Pacu and piranhas). In: Checklist of the Freshwater Fishes of South and Central America (Reis R.E., Kullander S.O. & Ferraris C.J. Jr., eds.). 2003, 182-196.
29. Reenamole GR, George D'cruz F. New Record of Exotic Fish Red Bellied Pacu, *Piaractus brachypomus* (Cuvier, 1818) From Vellayani Fresh Waterlake, Southwest Coast of India. International Journal of Science and Research (IJSR). 2014; 4(12):1106-1110.
30. Singh AK. Apprehensions and issues related to pacu *Piaractus brachypomus* (Cuvier 1818) farming in India. Journal of Fisheries Research. 2018; 2(1):19-23.
31. Singh A, Lakra WS. Risk and benefit assessment of alien fish species of the aquaculture and aquarium trade into India. Reviews in Aquaculture. 2011; 3(1):3-18
32. Unmesh Katwate, Deepak Apte, Rupesh Raut. Invasion in our Rivers. Hornbill. 2012, 42-45
33. Roshni K, Renjithkumar CR, Kurup BM. Record of a newly introduced fish, red-bellied pacu *Piaractus brachypomus* (Cuvier, 1818) (Characiformes, Serrasalmidae), in a tropical wetland system, India. Journal of Applied Ichthyology. 2014; 30(5):1037-1038.
34. Chattopadhyay NR, Kumari A, Sahoo U. Alien introduction and its impact on native fishery and aquatic biodiversity of West Bengal, India. Aquaculture Asia Magazine. 2011; 16(2):20-23
35. Laxmappa B. Exotic fish species in aquaculture and aquatic ecosystem in Telangana State, India. Journal of Aquatic Biology and Fisheries. 2016; 4:1-7.
36. Knight JM, Balasubramanian S. On a record of two alien fish species (Teleostei: Osphronemidae) from the natural waters of Chennai, Tamil Nadu, India. Journal of Threatened Taxa. 2015; 7(3):7044-7046.
37. Rick Spilman. Beware the Red-Bellied Pacu? Testicle Biting Fish Invading Denmark? (On-line). 2013
38. Ardura A, Pola IG, Genuino I, Gomes V, Garcia-Vazquez E. Application of barcoding to Amazonian commercial fish labelling. Food Research International. 2010; 43:1549-1552.
39. Buck EH. Seafood marketing: Combating fraud and deception. (Internet document. URL:<http://nationalaglawcenter.org/>). 2010am.
40. Cawthorn DM, Steinman HA, Witthuhn RC. DNA barcoding reveals a high incidence of fish species misrepresentation and substitution on the South African market. Food Research International. 2012; 46:30-40.
41. Cline E. Marketplace substitution of Atlantic salmon for Pacific salmon in Washington State detected by DNA barcoding. Food Research International. 2012; 45:388-393.
42. Galimberti A, De Mattia F, Losa A, Bruni I, Federici S, Casiraghi M et al. DNA barcoding as a new tool for food traceability. Food Research International. 2013; 50(1):55-63.
43. Marko PB, Lee SC, Rice AM, Gramling JM, Fitzhenry TM, McAlister JS. Mislabelling of a depleted reef fish. Nature. 2004; 430:309-310.
44. Ogden R. Fisheries forensics: the use of DNA tools for improving compliance, 306 traceability and enforcement in the fishing industry. Fish and fisheries. 2008; 9(4):462-472
45. Garcia-Vazquez E, Horreo JL, Campo D, Machado-Schiaffino G, Bista I, Triantafyllidis A. Mislabeling of two commercial North American hake 263 species suggests underreported exploitation of offshore hake. Transactions of the American Fisheries Society. 2009; 138:790-796.
46. Leonart J, Taconet M, Lamboeuf M. Integrating information on marine species identification for fishery purposes. Marine Ecology: Progress Series. 2006; 316:231-238.
47. Handy SM, Deeds JR, Ivanova NV, Hebert PDN, Hanner RH, Ormos A. A single-laboratory validated method for the generation of DNA barcodes for the identification of fish for regulatory compliance. Journal of AOAC International. 2011; 94:201-210.
48. Triantafyllidis A, Karaikou N, Perez J, Martinez JL, Roca A, Lopez B et al. Fish allergy risk derived from ambiguous vernacular fish names: Forensic DNA-based detection in Greek markets Food Research International. 2010; 43(8):2214-2216.
49. Smriti S, Islam T, Begum MK, Punom NJ, Mitra S, Hossain A et al. Fraudulence Detection in Fish Marketing of Bangladesh using DNA Barcoding. Journal of Bangladesh Academy of Sciences. 2017; 41(1):17-27.
50. Moravec F. Nematodes of freshwater fishes of the Neotropical Region. Academia, Publishing House of the Academy of Sciences of the Czech Republic. 1998, 464.
51. Froese, Pauly. *Piaractus brachypomus* Ecological Risk Screening Summary U.S. Fish and Wildlife Service, Web Version. <https://www.fws.gov/fisheries/>. 2010.
52. Robins CR, Bailey RM, Bond CE, Brooker JR, Lachner EA, Lea RN et al. World fishes important to North Americans. Exclusive of species from the continental waters of the United States and Canada. American Fisheries Society Special Publication. 1991, 21.
53. Caguan AG. Red-bellied Pacu in the Philippines. Journal of Environmental Science and Management. 2007; 10(1):42-47.
54. Singh AK. Emerging alien species in Indian aquaculture: prospects and threats. Journal of Aquatic Biology & Fisheries. 2014; 2(1):32-41.
55. de Brito MA, Schneider H, Sampaio I, Santos S. DNA barcoding reveals high substitution rate and mislabelling in croaker fillets (Sciaenidae) marketed in Brazil: The case of "pescada branca" (*Cynoscion leiacanthus* and *Plagioscion squamosissimus*). Food Research International. 2015; 70:40-46.
56. Deepak Jose, Harikrishnan M, Jenson Victor Rozario, Pradeep PJ, Saswata Maitra, Sameera S. Species misrepresentation revealed by molecular markers (COI rRNA): A reminder to the trade and export of *Macrobrachium rosenbergii* (submitted), 2019.